Synthesis, characterization and comparative thermal degradation study of Co(II), Ni(II) and Cu(II) complexes with Asparagine and Urea as mixed ligands
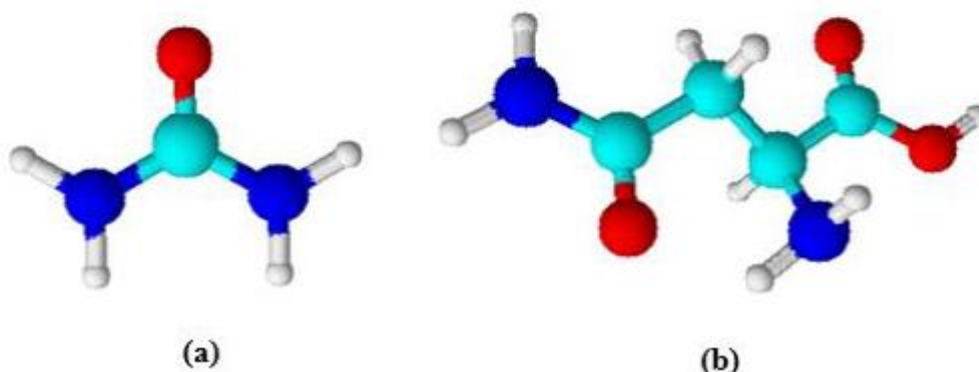

(a)

(b)

Figure 1. Structures of urea (a) and asparagine (b) molecules.

Ligands

Physicochemical and biological activity studies on complexes of some transition elements with mixed ligands of glycine and urea

Quality control

Simple, fast and inexpensive method for determination of ranitidine hydrochloride based on conductometric measurements

DNA breathing

Dark breather using symmetric morse, solvent and external potentials for DNA breathing

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Editorial

In the twilight of this year, the readers of Eclética Química Journal will find in this last issue interesting articles dealing with the synthesis and extensive physicochemical characterization of cobalt, nickel and copper complexes containing mixed ligands such as urea and glycine or urea and asparagine for which an octahedral structure was proposed. Antibacterial assays with *Bacillus* spp.*, Escherichia coli, Pseudomonas aeruginosa* and *Staphylococcus aureus* were performed for the metallic complexes containing urea and glycine, but only the nickel derivative showed some activity against *Escherichia coli.* In the sequence, the developing of a fast, simple and inexpensive alternative method for determining ranitidine in generic formulations without any sample pretreatment was described. It was based on conductometric titration of ranitidine hydrochloride by precipitation of AgCl using a solution of $AgNO_3$ as titrant. In another investigation, the dynamics and the quantum thermodynamics of DNA in Symmetric-Peyrard-Bishop-Dauxois model (S-PBD) with solvent and external potentials were analyzed and the transient conformational fluctuations using dark breather and the ground state wave function of the associate Schrodinger differential equation was described.

Assis Vicente Benedetti
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Eclética Química Journal

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SUMMARY

Synthesis, characterization and comparative thermal degradation study of Co(II), Ni(II) and Cu(II) complexes with Asparagine and Urea as mixed ligands

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ABSTRACT: New Co(II), Ni(II) and Cu(II) complexes with urea and asparagine as ligands have been synthesized in (M:L_1:L_2) molar ratio (where M= Co(II), Ni(II) and Cu(II), L_1 = urea, and L_2 = asparagine) then identified by micro analyses, molar conductance measurements, IR, ^{1}HNMR, Mass, UV-VIS spectroscopies and magnetic susceptibility measurements. Thermal degradation studies were carried out by thermal analysis. These complexes have the general formula $[M(L_1)(L_2)(H_2O)_n]Cl$. The molar conductance values in DMSO solvent show the electrolytic nature of these complexes, indicating the outer-sphere coordination of the chloride anions with metal ions. The three complexes have an octahedral structure with urea molecule showing two modes of coordination. Thermal analysis study shows the rapid decomposition reaction for Ni complex and the highest thermal stability for Cu complex. The kinetic parameters were determined from the thermal decomposition data using the Coats-Redfern method. Thermodynamic parameters were calculated using standard relations.
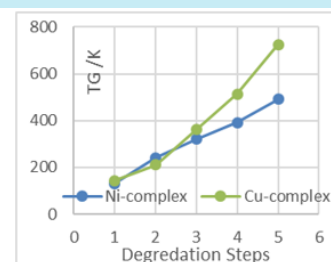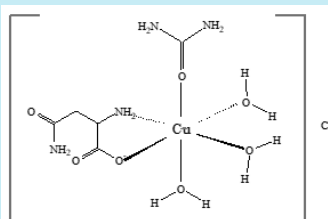
New complexes of urea and asparagine coordinated to M (Co^{2+}, Ni^{2+} and Cu^{2+}) were prepared and an octahedral structure was proposed. Cu complex presented higher thermal stability.

1. Introduction

Recently, there has been renewed attention in the preparation and studies of mixed ligand transition metal complexes[1,2] due to their new useful properties such as magnetic exchange, photoluminescence, nonlinear optical property, electrical conductivity and antimicrobial activity[3-5].

Mixed ligand complexes containing amino acid as co-ligand are potential biomimetic models for metal-protein interaction[6]. Research has shown significant progress in utilization of transition metal complexes as drugs to treat a lot of human diseases like carcinomas, infection control, anti-inflammatory, diabetes and neurological disorders[7].

Urea, carbamide or carbonyldiamide $CO(NH_2)_2$ (Figure 1a), which has a remarkable role in many biological processes in decomposition of proteins and amino acid catabolism, was discovered in 1828 by Wöhler when evaporating a solution containing a mixture of potassium isocyanate and ammonium sulphate[8].

The mode of urea bonding with metal ions seems to be dependent upon the type and nature of the metal, lead(II) coordinates to the nitrogen atom, whereas iron(III), zinc(II) and copper(II)

coordinate to the oxygen of urea[9]. Also, there are different types of coordination of urea in its complexes with rare-earth iodides and perchlorates[10].
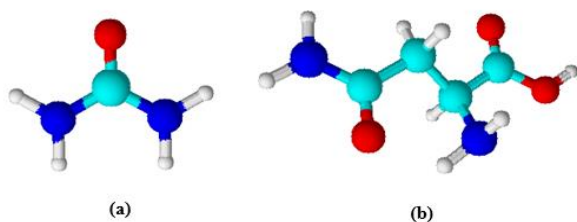

Figure 1. Structures of urea (a) and asparagine (b) molecules.

The amino acids are the main building units of all various forms of life and were earlier discovered as ingredient of natural products even before they were recognized as components of proteins[11]. The amino acid L-asparagine or 2-amino-3-carbamoylpropanoic acid (Fig. 1b) is a structural analog of L-aspartic acid. It was the first amino acid to be isolated from plants 200 years ago and because it has an N:C ratio of 2:4, this makes it an efficient molecule for the storage and transport of nitrogen in living organisms[12]. There are some similar thermal studies of various types of mixed ligands with transition metals[1, 2, 13-15], however, no previous studies on the synthesis, characterization and thermal studies of the mixed ligand complexes of urea and asparagine acid have been reported. Hence, the present work reports the preparation, characterization and thermal study of new mixed ligand complexes of urea and asparagine with Co(II), Ni(II) and Cu(II) ions.

2. Materials and methods

2.1 Chemicals

All chemicals such as solvents, metal(II) chlorides (i.e. $CoCl_2.6H_2O$, $NiCl_2.6H_2O$ and $CoCl_2.2H_2O$) were commercially available from BDH and were used without further purification.

2.2 Instrumentation

The melting points of the metal complexes were measured in glass capillary tubes with a Stuart Scientific Electrothermal melting point apparatus. TLC was carried out on silica gel GF_{254} plates (mn-kieselgel G., 0.2 mm thickness) with a 3:1 v/v ethyl acetate / petroleum ether solution as eluent mobile

at room temperature. The plates were scanned under ultraviolet light lamp of 254 nm. The CHN elemental analysis of the complexes was carried out by Vario ELFab. Chloride was determined volumetrically by silver nitrate. The amount of H_2O was determined gravimetrically using weight loss method. Perkin-Elmer 2380 flame atomic absorption spectrophotometer was used for the determination of metal content. Jenway conductivity meter model 4510 was used for measuring the molar conductance of the freshly prepared metal complexes solutions (10^{-3} mol L^{-1} in DMSO) at room temperature. IR spectra of the metal complexes were measured in the range 200-4000 cm^{-1} with a FT/IR–140 (Jasco, Japan). Varian FT-300 MHz spectrometer was used for recording ^{1}HNMR spectra in d_6DMSO solvent and TMS as internal standard. Mass spectra were recorded in a Jeol JMS600 spectrometer. The electronic spectra of the complexes were measured in the range 400–800 nm, using UV–VIS spectrophotometer Specord 200, Analytilk Jena (Germany). The magnetic susceptibility of the solid complexes was measured at room temperature using Gouy's method by a balance from Johnson Metthey and Sherwood model. The Differential Thermal Analysis (DTA) and Thermogravimetric Analysis (TGA) experiments were performed under nitrogen atmosphere using a platinum sample pan at a flow rate of 30 mL min^{-1} and a 10 °C min^{-1} heating rate for the temperature range 25–800 °C in Shimadzu DTA-50 and Shimadzu TGA-50H thermal analyzers, respectively, at Micro Analytical Center, Cairo University, Egypt.

2.3 Synthesis of mixed ligand complexes

Generally, the solid complexes were prepared by adding dropwise an ethanolic solution of hydrated metal(II) chlorides (0.01 mol) to an ethanolic solution of urea (0.01 mol) with stirring. The mixture was refluxed for 12 h with persistent stirring. A hot solution of 0.01 mol asparagine in 1:1 ethanol / water mixture ratio with drops of 1 mol L^{-1} NaOH was used to adjust the pH at 7-7.5 and to deprotonate NH_3^+ in the asparagine to NH_2. The mixture was refluxed for 2 h until the formation of colored precipitate occurred. All the solutions were in 1:1:1 molar ratio. The end products were filtered off and washed with distilled water to remove NaCl, followed by absolute ethanol until the solution became clear, and after

that the product was washed with DMF and left to dry[16]. The yield was 56%, 52% and 42% for Co, Ni and Cu-complexes, respectively.

3. Results and discussion

Complexes of Co(II), Ni(II) and Cu(II) with urea and asparagine are studied. Some physical properties, molar conductivity and analytical data are summarized in Tables 1 and 2. The elemental analysis proves that the complexes of Co(II), Ni(II) and Cu(II) with urea (ur) and asparagine (A_{asn}) ligands are of 1:1:1 (metal:ur:asn) molar ratio. The molar conductivity values indicate that the chloride anions are in the outer-sphere of these complexes.

Table 1. Some properties of the complexes.

Complex proposed formula	Color	M.p / °C	TLC No. of spots	TLC R_f	Molar conductivity Λ_m/S cm^2 mol^{-1}
[Co(ur)(asn)(H$_2$O)$_2$]Cl [Co(C$_5$H$_{15}$N$_4$O$_6$)]Cl	dark violet	203±1	One	0.18	133
[Ni(ur)(asn)(H$_2$O)$_2$]Cl [Ni(C$_5$H$_{15}$N$_4$O$_6$)]Cl	bluish green	185±1	One	0.24	128
[Cu(ur)(asn)(H$_2$O)$_3$]Cl [Cu(C$_5$H$_{17}$N$_4$O$_7$)]Cl	light violet	337±1	One	0.32	140

Table 2. The elemental analysis of the complexes.

Complex proposed formula	Molecular weight calc.	found	%C calc.	found	%H calc.	found	%N calc.	found	%M calc.	found	%Cl calc.	found
[Co(ur)(asn)(H$_2$O)$_2$]Cl [Co(C$_5$H$_{15}$N$_4$O$_6$)]Cl	321.58	321.61	18.67	18.67	4.70	4.70	17.42	17.42	18.33	18.32	11.03	11.04
[Ni(ur)(asn)(H$_2$O)$_2$]Cl [Ni(C$_5$H$_{15}$N$_4$O$_6$)]Cl	321.34	321.36	18.68	18.69	4.70	4.71	17.44	17.44	18.27	18.26	11.03	11.05
[Cu(ur)(asn)(H$_2$O)$_3$]Cl [Cu(C$_5$H$_{17}$N$_4$O$_7$)]Cl	344.21	344.23	17.44	17.45	4.98	4.89	16.28	16.28	18.46	18.46	10.29	10.31

3.1 IR Spectra of urea-asparagine complexes

In these complexes, urea acts in two ways: as a monodentate ligand through oxygen of C=O, or as a bidentate through nitrogen of two NH$_2$ groups, while asparagine acts as an anion bidentate molecule, through COO$^-$ group and NH$_2$ group. The assignment of the distinctive bands is summarized in Table 3 and the IR spectra of complexes are shown in Figures 2 to 4.

The IR spectra of the complexes show additional broad bands in the range 3386-3430 cm^{-1} due to the υ(OH) stretching of water molecule. Coordinated water is also identified by the appearance of ρ_r (rocking) and ρ_w (wagging) approximately at 875 cm^{-1} and 521 cm^{-1}, respectively. These results agree with the elemental analysis and thermogravimetric studies[17]. The υ(NH$_2$) stretching vibrations of free urea at υ_s3353 cm^{-1} and υ_{as}3466 cm^{-1} were shifted to lower wave numbers in the spectra of the complexes of Co(II) and Ni(II). This fact shows that the υ(NH$_2$) group must be involved in coordination while υ(CO) shifted to higher frequency[18].

Table 3. Main IR bands (cm^{-1}) of the urea-asparagine complexes.

Urea	Asparagine	[Co(ur)(asn)(H$_2$O)$_2$]Cl	[Ni(ur)(asn)(H$_2$O)$_2$]Cl	[Cu(ur)(asn)(H$_2$O)$_3$]Cl	Assignment
-	3110m	-	-	-	υ(NH$_3^+$)
3353m	3182m	ur-3320w asn-3185w,br asn-3227br	ur-3250 asn-3179br asn-3235br	ur-3298 m asn-3189br asn-3265 m	υ_S(NH$_2$)
3466 m	3182m	ur-3420br asn-3185w,br asn-3320w,br	ur-3430br asn-3179br asn-3235br	ur-3337m asn-3189br asn- 3386m	υ_{as}(NH$_2$) H$_2$O, υ(OH)
1618br	1644m	1637w	1637w	1629m	δ(NH)
-	1428s	1412 s	1420w	1423w	υ_s(COO$^-$)
-		1509 w	1509 w	1509w	υ_{as}(COO$^-$)
1695w	1745w 1681m	ur-1719w asn-1773w asn-1662m	ur-1725 s asn-1773w asn-1655m	ur-1629m asn-1773w asn- 1665m	υ(CO)
1468br	υ(C-N)1074s υ(O=C-N)1399m	ur-1446w asn-1081m asn-1409w,br	ur-1459m asn-1040 m asn-1410 w	ur-1490w asn-1039w asn1413m-	υ(C-N)
-	2874w	2873 w	2858 m	2857w	υ(CH$_2$)
-	1428s	1445w	1420w	1439w	δ(CH$_2$)
-		472 w	476w	454m	υ(M-O)
-		422w	421w	413m	υ(M-N)

s = strong,   m = medium,   br = broad,   w = weak,   w,br = weak and broad
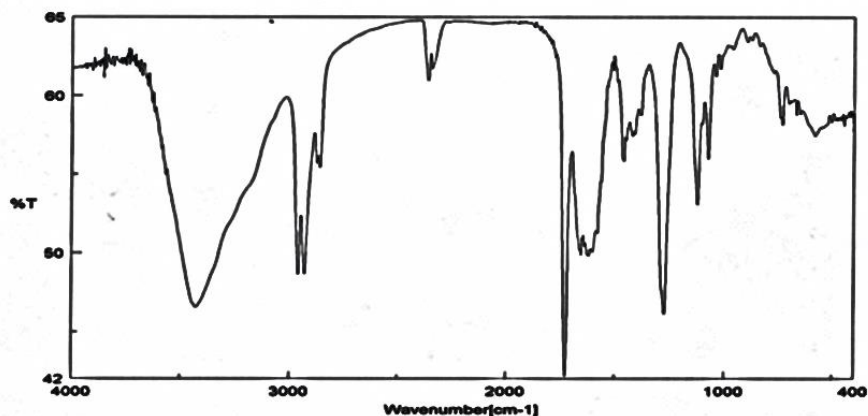

Figure 2. IR spectrum of [Co(ur)(asn)(H$_2$O)$_2$]Cl.

Figure 3. IR spectrum of [Ni(ur)(asn)(H$_2$O)$_2$]Cl.
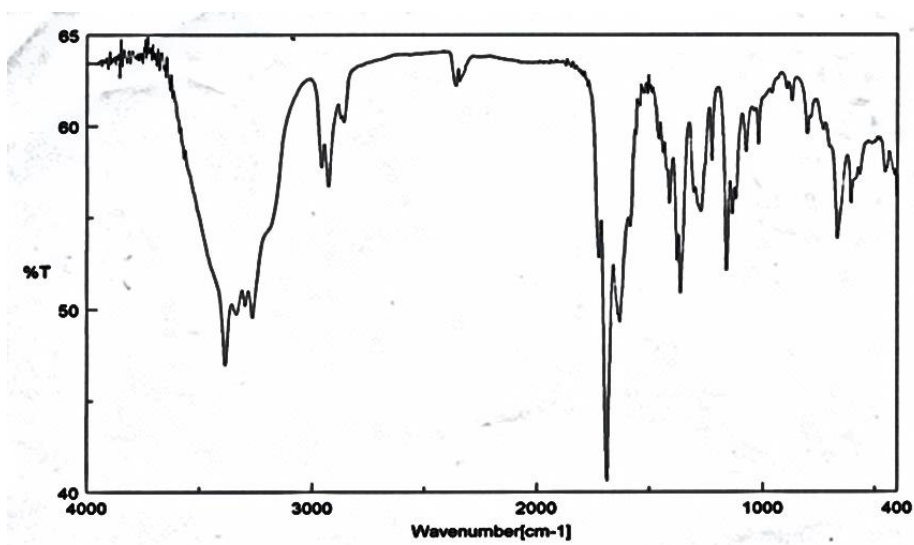
Figure 4. IR spectrum of [Cu(ur)(asn)(H$_2$O)$_3$]Cl.

The IR spectrum of the Cu(II)-complex showed a new band at 1629 cm^{-1} assigned to υ(C=O-Cu(II)) with slight change in υ(NH$_2$) vibration[19]. In comparison with asparagine, υ_s(COO$^-$) and υ_{as}(COO$^-$) shift to lower wave numbers, confirming the monodentate nature of the coordinated carboxylate group[20].

The υ(NH$_3^+$) band at 3110 cm^{-1}, which is specific for the zwitterion in asparagine, vanished in the spectra of the complexes after the deprotonation of NH$_3^+$ to NH$_2$. Therefore, the higher wave numbers shift of the bands assigned to υ_{as}(NH$_2$) and υ_s(NH$_2$) indicates that the NH$_2$ group is imminently involved in the coordination[20].

IR of the prepared complexes showed weak bands in the range of 476-454 cm^{-1} and 422-413 cm^{-1}, attributed to υ(M-O) and υ(M-N), respectively[21]. Other bands are listed in Table 3.

3.2 ^{1}HNMR spectra of urea-asparagine complexes

^{1}HNMR spectra of [Co(ur)(asn)(H$_2$O)$_2$]Cl, [Ni(ur)(asn)(H$_2$O)$_2$]Cl and [Cu(ur)(asn)(H$_2$O)$_3$]Cl complexes show various signals which were summarized in Table 4. Urea shows a new signal at 5 and 5.1 ppm in Co(II) and Ni(II) complexes, respectively, for the amide groups coordinated to the metal atom without proton displacement[22], while in Cu(II) complex only carbonyl group is coordinated to metal[22]. The signals at 3.2, 3.1 and 2.85 ppm are assigned to CH group, whereas signals at 2.9, 2.45 and 2.5 ppm of CH$_2$ group are observed for Co(II), Ni(II) and Cu(II) complexes, respectively. The appearance of a new signal around 2.6-2.7 ppm is attributed to NH$_2$ group of asparagine and the amide group shows signals in the range 6.3-6.7 ppm[23]. The coordinated H$_2$O shows a new signal around 3.5-3.7 ppm[24].

Table 4. ^{1}HNMR chemical shift of free urea and asparagine ligands and their complexes.

System	$(CH)_\alpha$	$(CH)_\beta$	NH_3^+	$NH_{2(asn)}$	$NH_{2(ur)}$	H_2O
Urea	-	-	-	-	6-7.5	-
Asparagine	4.6	2.6	7-8	6.9-7.6	-	-
$[Co(ur)(asn)(H_2O)_2]Cl$	3.2	2.9	-	6.6, 2.7	5 (bonding)	3.7
$[Ni(ur)(asn)(H_2O)_2]Cl$	3.1	2.45	-	6.3, 2.7	5.1 (bonding)	3.55
$[Cu(ur)(asn)(H_2O)_3]Cl$	2.85	2.5	-	6.7. 2.6	6.4 (nonbonding)	3.5

3.3 Mass spectra of urea – asparagine complexes

The mass spectra of Co(II), Ni(II) and Cu(II) complexes with urea and asparagine ligands exhibited the molecular ion peaks at m/z (calc. 321.58, found 321.61; calc. 321.34, found 321.36 and calc. 344.21, found 344.23), respectively.

The molecular ion of $[Co(ur)(asn)(H_2O)_2]Cl$ complex loses NH_4Cl and $H_2NCH_2COO^-$ leaving ions at m/z 268.13 and 247.07, respectively; then loses NH_2 and H_2O fragments, giving an ion at m/z 212.08. The spectrum of $[Ni(ur)(asn)(H_2O)_2]Cl$ complex shows a peak at m/z 267.89, indicating the loss of H_2O and $\frac{1}{2}Cl_2$. The molecule of $[Cu(ur)(asn)(H_2O)_3]Cl$ loses $H_2O + NH_2$ and $\frac{1}{2}Cl_2 + H_2O$, leaving ions at m/z 310.07 and 290.76, respectively. The ion at m/z 310.07 loses CO_2, leaving an ion at m/z 266.09, which further loses $\frac{1}{2}N_2$ to leave an ion at m/z 252.01. Afterwards, this last ion gives a new peak at m/z 234, indicating loss of H_2O and the remaining fragment loses another CO, leaving an ion at m/z 206.05.

3.4 Magnetic and electronic spectral studies

The electronic spectra of the Co(II), Ni(II) and Cu(II) complexes as well as their magnetic moment data have provided good evidence for the structures of these complexes as shown in Table 5. For $[Co(ur)(asn)(H_2O)_2]Cl$, hexa-coordination is suggested as in Figure 5a, based on the appearance of bands at 18248 cm^{-1} and at 14534 cm^{-1} (Figure 6), which were attributed to the $^4T_{1g} \rightarrow ^4T_{1g}(P)$ (υ_3) and $^4T_{1g} \rightarrow ^4A_{2g}$ (υ_2) transitions, respectively[25]. The third band, υ_1, could not be observed due to the limited range of the instrument used (200-1100 nm). Also, the magnetic moment of 4.81 B.M is within the range reported for a high-spin octahedral geometry around the Co(II) ion[26].

Table 5. Magnetic moments and electronic spectral data in DMSO solution for the complexes.

Complex	μ_{eff} / B.M	Charge transfer bands / cm[-1]	d-d transition bands / cm[-1]	Proposed structure
$[Co(ur)(asn)(H_2O)_2]Cl$	4.81	23585	18248, 14534	Octahedral
$[Ni(ur)(asn)(H_2O)_2]Cl$	2.9	24510	22727, 16181, 14619	Octahedral
$[Cu(ur)(asn)(H_2O)_3]Cl$	1.84	22727	16026	Distorted octahedral

$[Ni(ur)(asn)(H_2O)_2]Cl$ complex has a magnetic moment of 2.9 B.M, which is within the range reported for an octahedral geometry around the Ni(II) ion with a $^3A_{2g}$ ground term[27]. In addition, this complex has three bands in the UV-VIS spectrum (Figure 7): the band at 22727 cm^{-1} may be attributed to $^3A_{2g} \rightarrow ^3T_{1g}$ (υ_3); 16181 cm^{-1} due to $^3A_{2g} \rightarrow ^3T_{1g}$ (υ_2) and υ_1 at 14619 cm^{-1} in accordance with an

octahedral structure around the Ni(II) ion (Fig. 5a)[25, 28]. The electronic spectrum of [Cu(ur)(asn)(H$_2$O)$_3$]Cl (Fig. 5b and Fig. 8) shows a strong band at 16026 cm^{-1}. This band is due to $^2E_g \rightarrow {}^2T_{2g}$ transition and a distorted octahedral geometry is suggested[25]. The broadness of this band may be due to Jahn-Teller effect[25], which confirms the distorted octahedral geometry. The

magnetic moment value (1.84 B.M) is also within the range reported for the d^9-system containing one unpaired electron[29]. The bands at 23585 cm^{-1}, 24510 cm^{-1} and 22727 cm^{-1} should be attributed to the charge transfer transitions in the complexes [Co(ur)(asn)(H$_2$O)$_2$]Cl, [Ni(ur)(asn)(H$_2$O)$_2$]Cl and [Cu(ur)(asn)(H$_2$O)$_3$]Cl, respectively[30].

(a)

(b)

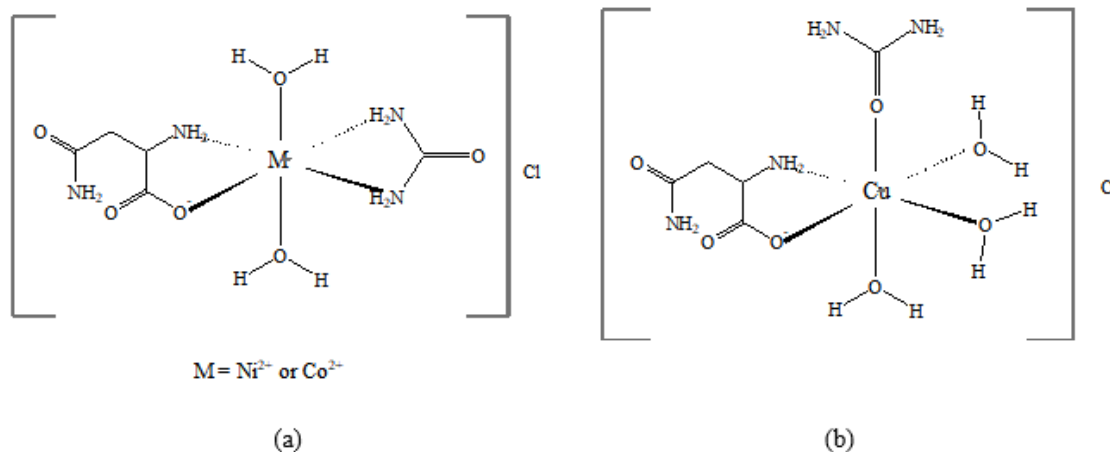M = Ni^{2+} or Co^{2+}

Figure 5. Suggested structure of the complexes.
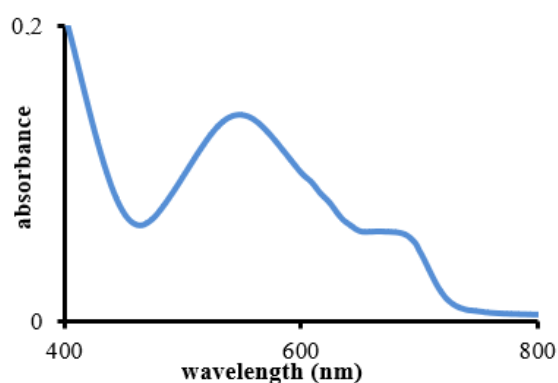

Figure 6. UV-VIS spectrum of [Co(ur)(asn)(H$_2$O)$_2$]Cl complex in DM.
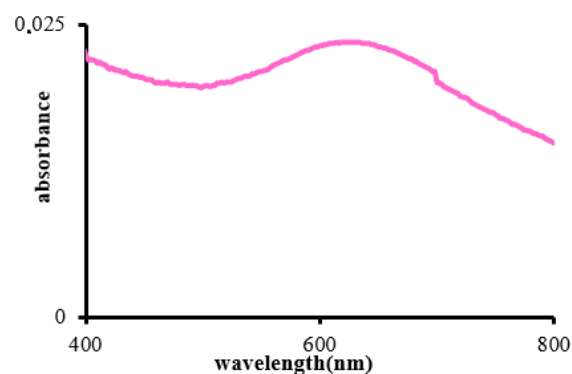
Figure 8. UV-VIS spectrum of [Cu(ur)(asn)(H$_2$O)$_3$]Cl complex in DMSO solution.
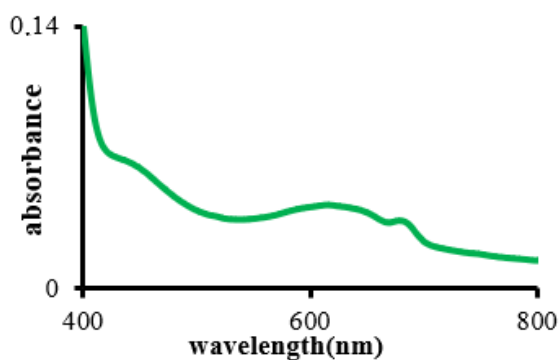
4.5 The Thermal degradation study

The TGA and DTA curves of the prepared Ni and Cu complexes are given in Figures 9 to 12. These curves characterize and compare the thermal degradation of these two complexes at 10 °C min^{-1} heating rate, under nitrogen and between 20-800 °C. For the evaluation of the thermal degradation kinetics parameters at a single heating rate (10 °C min^{-1}), the activation energy (E_a) and pre-exponential factor (Z) are determined by using the Coats-Redfen method for the reaction order $n \neq 1$. When the Coats-Redfern method is

Figure 7. UV-VIS spectrum of [Ni(ur)(asn)(H$_2$O)$_2$]Cl complex in DMSO solution.

linearized for a correctly-chosen order of reaction (*n*) yields the activation energy (*E*$_a$) from the slope of the equation:

$$\log\left[\frac{1-(1-\alpha)^{1-n}}{T^2(1-n)}\right]$$
$$= \log\left[\frac{ZR}{qE_a}\left(1-\frac{2RT}{E_a}\right)\right]$$
$$-\frac{E_a}{2.303RT}$$
$$for\ n \neq 1$$

where: α = fraction of weight loss, T = temperature (K), Z = pre-exponential factor, R = molar gas constant, q = heating rate and n = reaction order estimated by Horovitz-Metzger method.

The thermodynamic parameters of the thermal degradation step: enthalpy (ΔH^*), entropy (ΔS^*), and Gibbs energy (ΔG^*) of activation are calculated using the following standard equations:

$$\Delta S^* = R \ln \frac{Zh}{kT_{max}}$$
$$\Delta H^* = E_a - RT_{max}$$
$$\Delta G^* = \Delta H^* - T_{max}\Delta S^*$$

The characteristics of the thermal degradation of these two complexes recorded on the TG/DTG/DTA curves, their kinetics and thermodynamics parameters extracted from these curves are given in Tables 6-9.
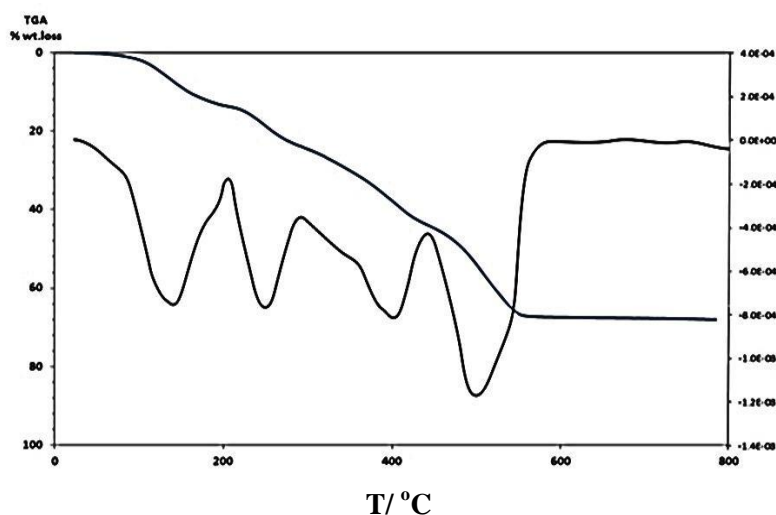

Figure 9. TG and DTG curves of [Ni(ur)(asn)(H$_2$O)$_2$]Cl complex.
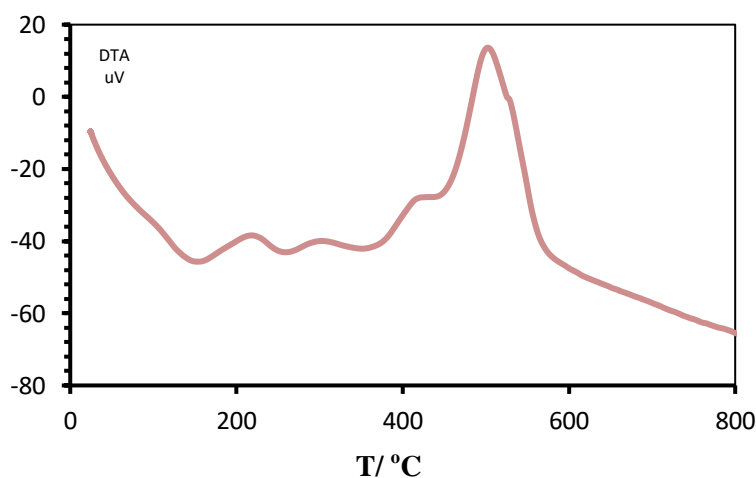
Figure 10. DTA curve of [Ni(ur)(asn)(H$_2$O)$_2$]Cl complex.
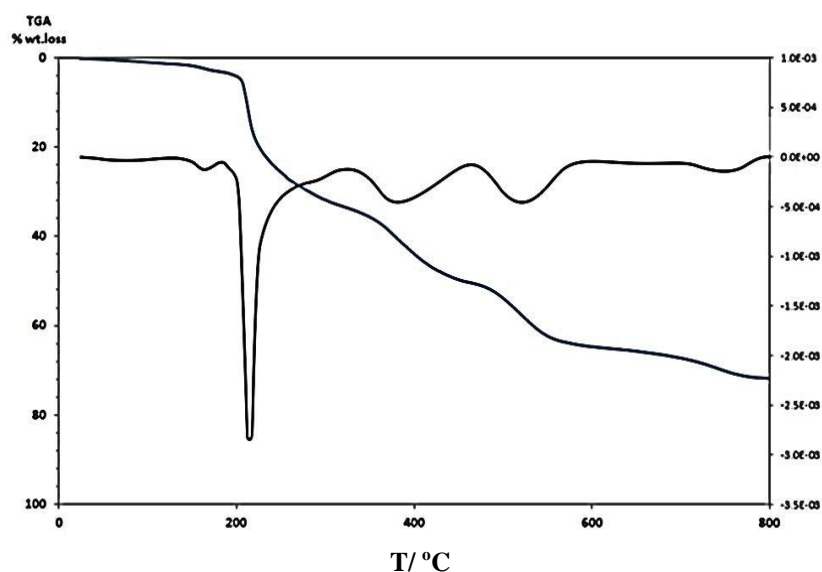
Figure 11. TG and DTG curves of [Cu(ur)(asn)(H$_2$O)$_3$]Cl complex.
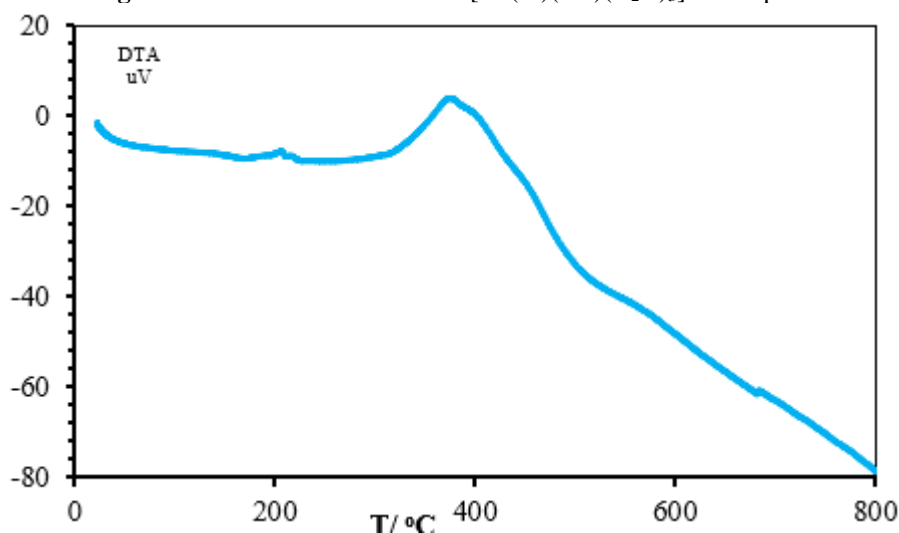
Figure 12. DTA curve of [Cu(ur)(asn)(H$_2$O)$_3$]Cl complex.

4.5.1 Thermal analysis of [Ni(ur)(asn)(H$_2$O)$_2$]Cl

The thermolysis of [Ni(ur)(asn)(H$_2$O)$_2$]Cl (Tables 6 and 7) and (Figures 9 and 10) involves several successive steps at 24-186, 186-274, 274-351, 351-428 and 428-552 °C. The first step represents the elimination of 100% coordinated H$_2$O molecules and 12.5% of a chloride atom (calc. 12.59%, found 12.58%) with activation energy E_a = 93 kJ mol^{-1} and a T_{DTG} peak at 132 °C. The second step corresponds to the loss of the remaining 87.5% Cl atom (calc. 12.12%, found 12.11%) which has E_a of 124 kJ mol^{-1} with T_{DTG} at 242 °C and exothermic peak T_{DTA} at 248 °C. According to the DTG curve (Figure 8), the third step is assigned to the removal of 42.86% urea

molecule (calc. 8.01%, found 8.00%) and with E_a and reaction order (*n*) of 114 kJ mol^{-1} and 0.1, respectively. The fourth step which corresponds to the loss of the remaining urea molecule and 3.52% of asparagine (calc. 12.12%, found 12.11%) has E_a = 129 kJ mol^{-1}. The final step corresponds to the 59.82% loss of asparagine (calc. 24.41%, found 24.44%) with E_a = 131 kJ mol^{-1} and a T_{DTG} peak at 491 °C. The final residue is NiO and carbon (2.67%) as ash (O=12.2%asn, C=24.46%asn) (calc. 33.21%, found 33.21%). The values of ΔS*, ΔH* and ΔG* are: -12.4, -117.9, -165, -120.4 and -144.5 J K^{-1} mol^{-1}; 91.9, 122, 111.3, 125.7 and 126.9 kJ mol^{-1}; and 93.5, 150.5, 164.3, 172.8 and 197.8 kJ mol^{-1}, respectively, for the steps observed in the thermal decomposition of the complex.

19

Table 6. Characteristic parameters of thermal decomposition (10 °C min⁻¹) for [Ni(ur)(asn)(H₂O)₂]Cl.

Comp.	Steps	TGA				DTA		mass loss
		Δm % found (calc.)	T_i/°C	T_f/°C	T_{DTG}	T_{DTA}	Heat	
[Ni(ur)(asn)(H₂O)₂]Cl	1	12.58 (12.59)	24	186	132	134	endo	-[100% H₂O+12.5% Cl]
	2	9.66 (9.66)	186	274	242	248	exo	-[87.5% Cl]
	3	8.00 (8.01)	274	351	321	321	exo	-[42.86% ur]
	4	12.11 (12.12)	351	428	391	408	exo	-[57.14% ur+3.52% asn]
	5	24.44 (24.41)	428	552	491	500	exo	[59.82% asn]
Final residue NiO +2.67C (O=12.2% asn, C=24.46% asn): 33.21% (33.21%)								

Table 7. Kinetic and thermodynamic parameters of the thermal decomposition of [Ni(ur)(asn)(H₂O)₂]Cl.

Comp.	Steps	r	n	Z / s⁻¹	T_{max} / K	E_a / kJ mol⁻¹	ΔS^* / J K⁻¹ mol⁻¹	ΔH^* / kJ mol⁻¹	ΔG^* / kJ mol⁻¹
[Ni(ur)(asn)(H₂O)₂]Cl	1	0.9794	2.6	6.2x10¹¹	132	93	-12.4	91.9	93.5
	2	0.9447	0.9	3.5x10⁶	242	124	-117.9	122	150.5
	3	0.9998	0.1	1.6x10⁴	321	114	-165	111.3	164.3
	4	0.9975	4.9	4.2x10⁶	391	129	-120.4	125.7	172.8
	5	0.9974	2.8	2.9x10⁵	491	131	-144.5	126.9	197.8

r = correlation coefficient of the linear plot, n = order of reaction, Z = pre-exponential factor.

Table 8. Characteristic parameters of thermal decomposition (10 °C min^{-1}) for [Cu(ur)(asn)(H$_2$O)$_3$]Cl.

Comp.	Steps	TGA				DTA		mass loss
		Δm % found (calc.)	T$_i$/°C	T$_f$/°C	T$_{DTG}$	T$_{DTA}$	Heat	
[Cu(ur)(asn)(H$_2$O)$_3$]Cl	1	2.98 (2.99)	22	178	143	168	endo	-[19.05% H$_2$O]
	2	29.35 (29.35)	178	306	211	207	exo	-[80.95%H$_2$O+100%Cl + 36.36% ur]
	3	17.35 (17.37)	306	449	362	376	exo	-[63.64% ur+16.46% asn]
	4	16.44 (16.46)	449	666	514	-	-	-[43.20% asn]
	5	5.51 (5.49)	666	785	725	-	-	-[14.40% asn]

Final residue CuO + 1.5% C (O=12.2% asn, C=13.74% asn): 28.37% (28.34%)

Table 9. Kinetic and thermodynamic parameters of the thermal decomposition of [Cu(ur)(asn)(H$_2$O)$_3$]Cl.

Comp.	Steps	r	n	Zvz / s^{-1}	T$_{max}$/ K	E$_a$ / kJ mol^{-1}	ΔS* / J K^{-1} mol^{-1}	ΔH* / kJ mol^{-1}	ΔG* / kJ mol^{-1}
[Cu (ur)(asn)(H$_2$O)$_3$]Cl	1	0.9916	4.8	2.4x10^8	143	98	-192.6	96.8	124.3
	2	0.9689	5	1.9x10^7	211	119	-102.7	117.2	138.7
	3	0.9855	2.2	3.8x10^8	362	134	-82.3	135	165.2
	4	0.9946	3.5	2.7x10^6	514	123	-126.3	118.7	183.6
	5	0.9972	4.9	3.2x10^5	725	137	-146.9	134	240.5

r = correlation coefficient of the linear plot, n = order of reaction, Z = pre-exponential factor.

4.5.2 Thermal analysis of [Cu(ur)(asn)(H$_2$O)$_3$]Cl

The TG and DTG curves of [Cu(ur)(asn)(H$_2$O)$_3$]Cl (Tables 8 and 9) and (Figures 11 and 12) show five steps of a continuous mass loss with DTG peaks indicating slow mass losses. The first step (22-178 °C) at T$_{DTG}$ 143 °C is attributed to the release of 19.05% of coordinated H$_2$O (calc. 2.99%, found 2.98%). The remaining loss of H$_2$O, and the loss of 100% Cl and 36.36% of urea occur in the second step (178-306 °C). Third (306-449 °C), fourth (449-666 °C) and fifth (666-785 °C) steps are due to the release of [63.64%ur+16.46%asn], [43.20%asn] and [14.40%asn] fragments (calc. 17.37%, found

17.35%; calc. 16.46%, found 16.44% and calc. 5.49%, found 5.51%), respectively, at the T$_{DTG}$ peaks at 362, 514 and 725 °C (Figure 9), respectively. The E$_a$ calculated of these five steps are 98, 119, 134, 123 and 137 kJ mol^{-1}, respectively, and the values of ΔS*, ΔH* and ΔG* are: -192.6, -102.7, -82.3, -126.3 and -146.9 J K^{-1} mol^{-1}; 96.8, 117.2, 135.0, 118.7 and 134.0 kJ mol^{-1}, and 124.3, 138.7, 165.2, 183.6 and 240.5 kJ mol^{-1}, respectively, for the steps observed in the thermal decomposition of the complex.

4.5.2 General remarks of thermal degradation:

1.   Thermal analysis confirms the presence of coordinated water molecules.

2. Sharp peak in Ni-complex means that the leaving parts move away faster than that in Cu-complex.

3. The releasing of the urea ligand before asparagine ligand may be due to non-ionic bonding of this ligand with the metal ions.

4. The first step which represents the dehydration of coordinated water is faster in Ni complex (E_a = 93 kJ mol^{-1}) than in Cu complex (E_a = 98 kJ mol^{-1}) (Figure 11a). The Cu complex is more stable than Ni complex (Figure 11b), which is indicated by the higher T_{max} value.

5. The chloride evolution starts at the first step in Ni complex and in the second step the Cu complex, in accordance with literature[31]. On the other side, the high value of T_{DTG} (211 °C) of Cu complex reflects its higher stability compared to Ni complex of T_{DTG} (132 °C).

6. Ni complex has a faster complete decomposition of its backbone (E_a = 131 kJ mol^{-1}) than Cu complex (E_a = 137 kJ mol^{-1}), making clear the higher stability of Cu complex.

7. The values of ΔG^* for a given complex, generally, increase significantly for the subsequent decomposition steps, consequence of the increase of $T\Delta S^*$ values from one step to another which exceed the ΔH^* values.

4. Conclusions

In this paper, some new complexes containing urea and asparagine ligands were prepared and characterized. The complexes have the following molecular formulae: $[M(L_1)(L_2)(H_2O)_n]Cl$ where M = Co(II), Ni(II) and Cu(II), L_1 = urea, and L_2 = asparagine. These complexes were characterized by elemental analysis, conductance measurements, IR, ^{1}HNMR and mass spectroscopy. Electronic spectra and magnetic measurements suggested an octahedral geometry for the complexes. Thermal analysis study showed faster decomposition reactions of the Ni complex and higher thermal stability of Cu complex.

5. Acknowledgments

6. References

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Physicochemical and biological activity studies on complexes of some transition elements with mixed ligands of glycine and urea

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ABSTRACT: The reaction of urea (ur) and glycine (gly) with the metal ions Co(II), Ni(II) and Cu(II) in ethanolic solution of $1M:1L_1:1L_2$ molar ratio (where M= Co(II), Ni(II) and Cu(II), and L_1 = urea L_2 = glycine) led to the preparation of complexes of the general formula $[M(ur)(gly)(H_2O)_2]Cl$. Elemental microanalysis (CHN), molar conductivity measurements, IR,^{1}HNMR, Mass and UV-VIS spectroscopic, and magnetic susceptibility measurements were used for the characterization of the compounds. Thermal analyses were used for the complexes degradation characterization. The complexes have an octahedral geometry and are of electrolytic nature in DMSO solvent with the absence of inner-sphere coordination of the chloride anion. An inhibition zone was observed for Ni-urea-glycine complex against *Escherichia coli* when the biological activity was considered.

An octahedral geometry is suggested for the complexes. An inhibition zone was observed for Ni-urea-glycine complex against *Escherichia coli*.

$M=Co^{2+}, Ni^{2+}, Cu^{2+}$
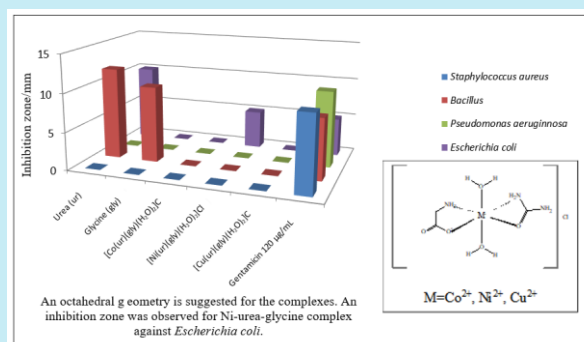
1. Introduction

The synthesis and study of mixed ligand transition metal complexes have been of growing interest[1, 2]. New materials with useful properties such as electrical conductivity photoluminescence, magnetic exchange, nonlinear optical property and antimicrobial activity can be provided by using mixed ligand transition metal complexes[3-5].

Urea $(CO(NH_2)_2)$ plays an important role in many biological processes such as decomposition of proteins and amino acids catabolism. In 1828, Wöhler discovered urea, when organic materials were prepared from inorganic substances.

All living things contain building blocks of amino acids[6], which were first discovered as constituents of natural products and then observed to be the major components of proteins.

All life forms on earth consist of the simplest proteinaceous amino acid, called glycine or amino acetic acid[7]. Glycine is a neutral, aliphatic, optically inactive nonessential amino acid[8] and it is the only protein amino acid that does not have optical isomers[9]. Most of the metal ions form mono, bis and tris complexes with glycine that acts as a bidentate ligand forming stable 5-membered chelating rings via the N atom of the amino group and O atom of carboxylate group[10].

The mixed ligand complexes of urea and glycine acid with Co(II), Ni(II) and Cu(II) ions were synthesized, characterized and thermally studied for the first time in this work.

2. Materials and methods

2.1 Materials

All Chemical reagents used were purchased from BDH and used as provided.

2.2 Synthesis of the complexes

Generally, the solid complexes were prepared by the same methodology previously described[11]. Briefly, an ethanolic solution of hydrated metal chloride (0.01 mol) was dropwise added in an ethanolic solution of the first ligand (urea 0.01 mol) with stirring. The mixture was refluxed for 12 h with constant stirring. A hot solution of 0.01 mol glycine in 1:1 ethanol / water mixture ratio was dropwise added to the urea / metal mixture and drops of 1 mol L^{-1} NaOH solution were used to adjust pH 7.0 - 7.5 to deprotonate NH_3^+ of the glycine to NH_2. The mixture was refluxed for 2 h until resulting in the formation of a colored precipitate. The resulting product was filtered off and then washed with distilled water to remove NaCl. The product was further washed with absolute ethanol/dimethylformamide (DMF) and left to dry. Acceptable yield percentage was obtained (52-66%).

2.3 Instrumentation

Glass capillary tubes were used to measure the melting points of the metal complexes in degrees celsius on a Stuart Scientific electrothermal melting point apparatus. Silica Gel GF_{254} plates (mn-kieselgel G., 0.2 mm thickness) was used for TLC. Vario ELFab instrument was used for elemental analysis (carbon, hydrogen and nitrogen) of complexes. Chloride was volumetrically or gravimetrically determined by silver nitrate. The amount of water was determined gravimetrically using weight loss method and also from thermal analysis. Perkin-Elmer 2380 flame atomic absorption spectrophotometer was used to measure the metal content. Jenway conductivity meter model 4510 was used to measure the molar conductance of 10^{-3} mol L^{-1} solutions of the metal complexes in dimethylsulfoxide (DMSO) solvent. IR spectra of the metal complexes were measured by using FT/IR-140 (Jasco, Japan). A Varian FT-300 MHz spectrometer in d_6-DMSO solvent was used for obtaining proton ^{1}HNMR spectra, using TMS as internal standard. Mass spectra were recorded on a JEOL JMS600 spectrometer. The electronic spectra of the complexes were measured in the range 400-800 nm using an UV–VIS spectrophotometer Specord 200, Analytilk Jena (Germany). The mass susceptibility (χ_g) of the solid complexes was measured at room temperature using Gouy's method on a magnetic susceptibility balance from Johnson Metthey and Sherwood model. Differential Thermal Analysis (DTA) and Thermogravimetric Analysis (TGA) were performed using the Shimadzu DTA-50 and Shimadzu TGA-50H thermal analyzers. The experiments were carried out in the temprature range from 25 to 800 ^{o}C under nitrogen atmosphere in a platinum pan, heating rate of 10 ^{o}C / min and flow rate of 30 mL min^{-1}. The antibacterial activity against four species of bacteria (*Staphylococcus aureus*, *Bacillus* spp., *Escherichia coli* and *Pseudomonas aeruginosa*) was tested by agar diffusion method. 1000 µg mL^{-1} concentration for each of these compounds were individually prepared in DMSO, then the filter paper disc (whatman No.1.5 mm diameter) was saturated with the solution of these compounds. The discs were placed on the surface of Millar Hinton agar dishes seeded with the strains of bacteria. The inhibition zones (mm) were measured after 24 h at 37 ^{o}C. DMSO and gentamicin (120 µg mL^{-1}) were used as control and reference, respectively.

3. Results and discussion

Complexes of Co(II), Ni(II) and Cu(II) with urea (ur) and glycine (gly) ligands have been prepared and characterized. Analytical data, physical properties, molar conductivity, and composition of the synthesized complexes are given in Tables 1 and 2. The molar conductivity values (135-149 S cm^2 mol^{-1}) reflect the electrolytic properties of these complexes. The single spot appearance in the TLC proves the purity of these complexes.

Table 1. Some physical properties of the complexes.

| Complex Proposed Formula | Color | M.p /°C | TLC | | molar conductivity Λ_m / S cm^2 mol^{-1} |
			No.of spots	Rf	
[Co(ur)(gly)(H$_2$O)$_2$]Cl [Co(C$_3$H$_{12}$N$_3$O$_5$)]Cl	dark violet	>350	One	0.28	149
[Ni(ur)(gly)(H$_2$O)$_2$]Cl [Ni(C$_3$H$_{12}$N$_3$O$_5$)]Cl	pale green	226±1	One	0.39	135
[Cu(ur)(gly)(H$_2$O)$_2$] Cl [Cu(C$_3$H$_{12}$N$_3$O$_5$)]Cl	light blue	>350	One	0.39	148

Table 2. Elemental analysis of the complexes.

| Complex proposed formula | Molar mass | | Elemental analysis | | | | | | | | | |
| | calc. | found | %C | | %H | | %N | | %M | | %Cl | |
			calc.	found	calc.	found	calc.	found	calc.	found	calc.	found
[Co(ur)(gly)(H$_2$O)$_2$]Cl [Co(C$_3$H$_{12}$N$_3$O$_5$)]Cl	264.53	264.03	13.62	13.65	4.57	4.58	15.89	15.91	22.28	22.32	13.40	13.45
[Ni(ur)(gly)(H$_2$O)$_2$]Cl [Ni(C$_3$H$_{12}$N$_3$O$_5$)]Cl	264.29	264.34	13.63	13.63	4.58	4.58	15.90	15.90	22.21	22.20	13.41	13.43
[Cu(ur)(gly)(H$_2$O)$_2$]Cl [Cu(C$_3$H$_{12}$N$_3$O$_5$)]Cl	269.14	269.19	13.38	13.39	4.49	4.49	15.61	15.61	23.61	23.61	13.17	13.17

3.1 IR spectra of urea - glycine complexes

The coordination sites of urea and glycine ligands in their complexes were investigated. The infrared spectra show that urea acts as a neutral bidentate ligand through C=O and NH$_2$ groups while glycine behaves as a bidentate anion ligand through COO$^-$ and NH$_2$ groups. IR spectra of urea-glycine complexes are represented in Figures 1, 2 and 3. Assignments of the characteristic bands are summarized in Table 3. As it was postulated, the metal complexes were quite different when compared with the free ligands.
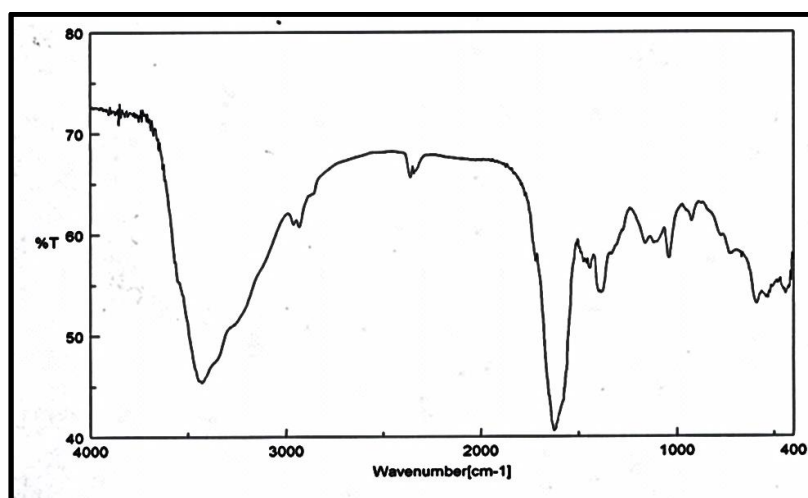
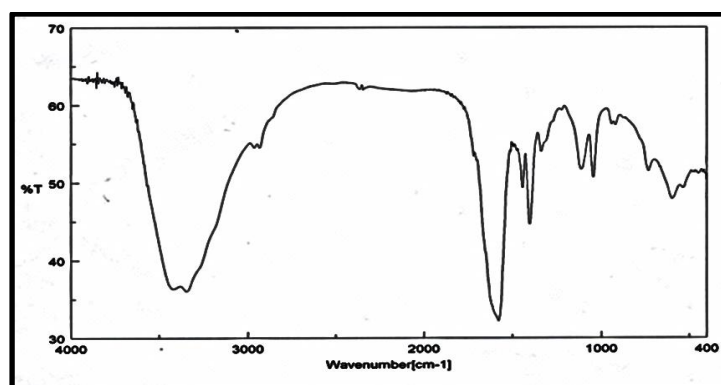**Figure 1.** IR spectrum of [Co(ur)(gly)(H$_2$O)$_2$]Cl complex.
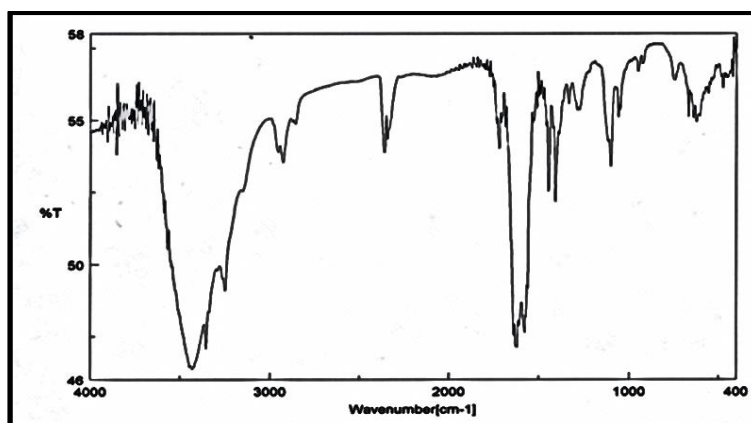
Figure 2. IR spectrum of [Ni(ur)(gly)(H$_2$O)$_2$]Cl complex.

Figure 3. IR spectrum of [Cu(ur)(gly)(H$_2$O)$_2$]Cl complex.

Table 3. Main IR bands (cm^{-1}) of the urea-glycine complexes

Urea	Glycine	[Co(ur)(gly)(H$_2$O)$_2$]Cl	[Ni(ur)(gly)(H$_2$O)$_2$]Cl	[Cu,(ur)(gly)(H$_2$O)$_2$]Cl	Assignment
-	3164-3007br	-	-	-	υ(NH$_3$$^+$)
3353m	-	ur-3250br gly-3185w,br	ur-3270br gly -3168w,br	ur -3248br gly -3160br	υ_s(NH$_2$)
3466 m	-	ur, gly - 3376br	ur,gly − 3346m	ur - 3355 m gly -3290w	υ_{as}(NH$_2$)
1618br	-	-	-	1609w	δ (NH)
-	1410m	1397 m	1402s	1408s	υ_s(COO$^-$)
-	1598br	1509w	1578s	1578m	υ_{as}(COO$^-$)
1695w	1715w	ur − 1625s gly −1718m	ur − 1680w gly −1718w	ur - 1618 m gly−1719m	υ(CO)
1468br	1033s	ur − 1474w gly -1040m	ur − 1475w gly -1045m	ur−1475w gly −1050 w	υ(CN)
-	2892br	2885br	2863 w.br	2857br	υ(CH$_2$)
-	1442m	1439m	1441s	1446s	δ (CH$_2$)
-	-	3431br	3422br	3432br	H$_2$O υ(OH)
-	-	451w	483 w	475 w	υ(M-O)
-	-	440w	460 w	421 w	υ(M-N)

s = strong, m = medium, br = broad, w = weak, w,br = weak and broad

The infrared spectral data of the complexes are as follows:

(1) All the complexes spectra show a broad band at 3422-3432 cm^{-1} that corresponds to the stretching mode of water existing in the complexes as identified by thermal and elemental analysis. The coordinated water is identified by the appearance of ρ_r (rocking) and ρ_w (wagging) at 925 cm^{-1} and 511 cm^{-1}, respectively[12].

(2) The amino groups of urea show lower-shift of 123-103 cm^{-1} and of 120-90 cm^{-1} for symmetrical and asymmetrical stretching υ(NH$_2$) frequencies, respectively. This strongly suggests that the nitrogen atom of amino group must be involved in complexation, and the appearance of a new band in the range of 406-460 cm^{-1}, assigned to υ(M-N) vibration, confirms this proposition[13, 14].

(3) A new band at 1680-1618 cm^{-1} is attributed to υ(CO) from urea, assigned to υ(C=O–M).

(4) The characteristic bands in complexes spectra occur in the ranges 3185-3160 cm^{-1} and 3376-3290 cm^{-1} for symmetrical and asymmetrical υ(NH$_2$) group of glycine, respectively, which appears at lower wave number than the free υ(NH$_2$). Hence, coordination through nitrogen of the amino group is involved[15].

(5) The symmetrical υ(COO$^-$) and asymmetrical υ(COO$^-$) vibrations of glycine shift by 13-8 cm^{-1} and 89-20 cm^{-1}, respectively. This confirms that carboxylate is acting as a monodentate group[16]. Glycine acts as monobasic bidentate, through the nitrogen of amino and oxygen of carboxylate groups in these complexes[17, 18].

(6) The IR spectra in the range 483-451cm^{-1} and 460-421cm^{-1} show bands of low intensity due to stretching vibrations of υ(M-O) and υ(M-N), respectively[13, 14].

3.2. ^{1}HNMR spectra of urea-glycine complexes

Complexes were investigated by using ^{1}HNMR spectra in d$_6$-DMSO and TMS (tetramethyl silane) as standard and data are in Table 4. [Co(ur)(gly)(H$_2$O)$_2$]Cl, [Ni(ur)(gly)(H$_2$O)$_2$]Cl and [Cu(ur)(gly)(H$_2$O)$_2$]Cl complexes show signals in the range 5.4-7.2 ppm attributed to the amide group of urea[19, 20]. The methylene group of glycine (-CH$_2$-) in Co(II), Ni(II) and Cu(II) complexes absorbs near 3.2, 3.2 and 3.1 ppm, respectively. NH$_2$ group shows signals at 2.9, 2.5 and 2.6 ppm, respectively[21, 22]. In urea, one amine and the carbonyl groups are coordinated to the central metal ion without displacement of NH$_2$ proton,

while in glycine presents a new signal in the range 2.5-2.9 ppm because of the deprotonation of NH^{3+} to NH_2. The appearance of a new signal around 3.5- 3.8 ppm confirms the presence of water molecules in the complexes[23].

Table 4. ^{1}HNMR chemical shift (ppm) of free urea and glycine ligands and of complexes.

System	$(CH)_\alpha$ α =Alpha	NH_3^+	$NH_{2(gly)}$	$NH_{2(ur)}$	H_2O
urea	-	-	-	6-7.5	-
glycine	3.5	8-7	-	-	-
$[Co(ur)(gly)(H_2O)_2]Cl$	3.2	-	2.9	5.4$_{(bonding)}$ 6.4$_{(nonbonding)}$	3.8
$[Ni(ur)(gly)(H_2O)_2]Cl$	3.2	-	2.5	4.7$_{(bonding)}$ 7.2$_{(nonbonding)}$	3.7
$[Cu(ur)(gly)(H_2O)_2]Cl$	3.1	-	2.6	5.5$_{(bonding)}$ 7$_{(nonbonding)}$	3.5

3.3 Mass spectra of urea-glycine complexes

The mass spectra of Co(II), Ni(II) and Cu(II) complexes with urea and glycine reveal molecular ion peaks at m/z (calc. 264.53, found 264.03 (4%)), (calc. 264.29, found 264.34 (11%)) and (calc. 269.14, found 269.19 (9%)), respectively.

The molecular ion of $[Co(ur)(gly)(H_2O)_2]Cl$ complex loses glycinate ($NH_2CH_2COO^-$) ion and $2H_2$ leaving an ion at m/z 185.67, which by its turn, loses H_2O, Cl, CO, NH_3 and H_2 giving an ion at m/z 85.01.
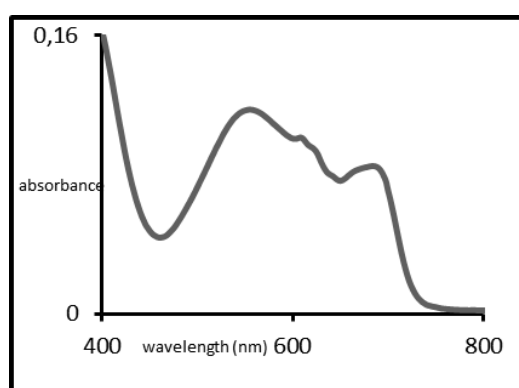
The mass spectrum of $[Ni(ur)(gly)(H_2O)_2]Cl$ complex exhibited a peak at m/z 244.90, indicating the loss of H_2 and NH_3, then this molecular ion loses H_2O and $\frac{1}{2}Cl_2$ leaving an ion at m/z 192.83, which further loses one more $H_2NCH_2COO^-$ affording an ion at m/z 118.87. The complex $[Cu(ur)(gly)(H_2O)_2]Cl$ loses [CO, $\frac{1}{2}Cl_2$] and H_2O to give ions at m/z 205.68 and 251.19, respectively.

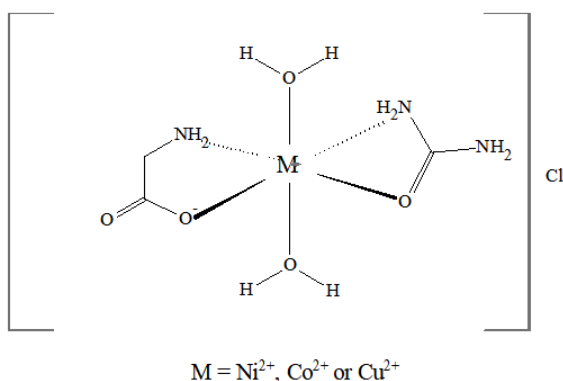*3.4 Electronic and magnetic spectral analysis*

The magnetic moments of the Co(II), Ni(II) and Cu(II) complexes as well as their electronic spectra data have provided good evidence for the structures of these complexes as shown in Table 5. $[Co(ur)(gly)(H_2O)_2]Cl$ hexa-coordination is suggested. This is based on the spectrum (Figure 5) recorded in DMSO solution which shows bands at 17985 cm^{-1} and 14482 cm^{-1}, due to transition of $^4T_{1g} \rightarrow {}^4T_{1g}(P)$ (υ3) and $^4T_{1g} \rightarrow {}^4A_{2g}$ (υ2), respectively[23]. The third band of the spectrum, assigned to υ1, could not be observed due to the limited range of the used instrument (200-1100 nm). $[Co(ur)(gly)(H_2O)_2]Cl$ has a magnetic moments of 4.76 B.M; this value is due to a high-spin octahedral geometry around the Co(II) ion as reported previously[24]. Moreover, the violet colour of octahedral Co(II) complexes is in good agreement with those previously reported[25].

Table 5. Magnetic moment and electronic spectral data in DMSO solution for the complexes.

complex	μ_{eff}/ B.M	charge transfer bands / cm^{-1}	d-d transition bands / cm^{-1}	proposed structure
[Co(ur)(gly)(H$_2$O)$_2$]Cl	4.76	23697	17985, 14492	octahedral
[Ni(ur)(gly)(H$_2$O)$_2$]Cl	3.2	23419	21459, 14970, 13477	octahedral
[Cu(ur)(gly)(H$_2$O)$_2$]Cl	1.43	24272	12987	distorted octahedral

Figure 4. UV-VIS spectrum of [Co(ur)(gly)(H$_2$O$_2$]Cl complex in the MSO solution.

From the above discussion (Figure 5) of [Co(ur)(gly)(H$_2$O$_2$]Cl can be suggested. Furthermore, previous studies proved that the broad bands centred at 23697 cm^{-1} should be assigned to charge-transfer transitions in [Co(ur)(gly)(H$_2$O)$_2$]Cl[26].

M = Ni^{2+}, Co^{2+} or Cu^{2+}

Figure 5. Suggested structure for the complex.

The magnetic moment data as well as the electronic spectrum data of the nickel complex are given in Table 5. The complex

[Ni(ur)(gly)(H$_2$O)$_2$]Cl has a magnetic moment value of 3.2 B.M consistent with an octahedral geometry around the Ni(II) ion with a $^3A_{2g}$ ground term, which lies in the range reported in the literature[27]. In addition, the complex has three bands in the UV-VIS recorded in DMSO solution (Figure 6): 21459 cm^{-1} may be due to the $^3A_{2g} \rightarrow ^3T_{1g}$ ($\upsilon3$); 14970 cm^{-1} due to $^3A_{2g} \rightarrow ^3T_{1g}$ ($\upsilon2$); 13477 cm^{-1} in the transition range of an octahedral structure around the Ni(II) ion ($\upsilon1$) (Figure 5)[28]. The green colour is also an additional evidence for the octahedral structure[26]. The band at 23419 cm^{-1} may be attributed to the charge transfer transition of [Ni(ur)(gly)(H$_2$O)$_2$]Cl complex[23].

[Cu(ur)(gly)(H$_2$O)$_2$]Cl (structure in Figure 5) has an electronic spectrum (Figure 7) that shows a strong band at 12987 cm^{-1} due to $^2E_g \rightarrow ^2T_{2g}$ transition, suggesting a distorted octahedral geometry[26]. The broadness in the band may be due to Jahn-Teller effect[29] and the proposed geometry is also supported by the blue colour of this complex[27]. The magnetic moment value of this complex 1.43 B.M agrees with the d^9 system containing one unpaired electron[26]. The observed band at 24272 cm^{-1} in the spectrum of the complex may be due to LMCT (L→M charge transfer transition) of [Cu(ur)(gly)(H$_2$O)$_2$]Cl complex[30].
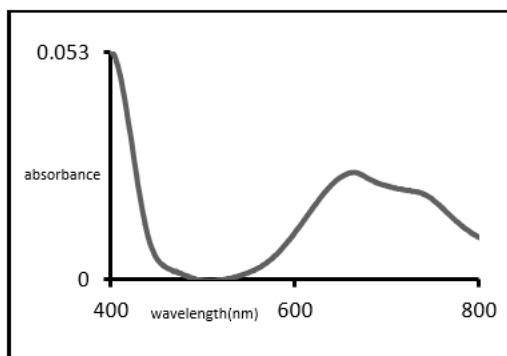
Figure 6. UV-VIS spectrum of [Ni(ur)(gly)(H$_2$O)$_2$]Cl complex in DMSO solution.

Figure 7. UV-VIS spectrum of [Cu(ur)(gly)(H$_2$O)$_2$]Cl complex in DMSO solution.

3.5. Thermal analysis of Cu-urea-glycine complex

The thermal and kinetic parameters for each step in the decomposition sequences of the Cu-complex were determined by using the integral Coast-Redfern equation. The Coats-Redfern method is linearized for a correctly-chosen order of reaction (n) and the activation energy (E_a) is obtained from the slope of the log [y] *versus* T^{-1} plot from Equation:

$$log \left[\frac{1-(1-\alpha)^{1-n}}{T^2(1-n)} \right]$$
$$= log \left[\frac{ZR}{qE_a} \left(1 - \frac{2RT}{E_a} \right) \right]$$
$$- \frac{E_a}{2.303RT} \; for \; n \neq 1 \quad \rightarrow 1$$

where: α = fraction of mass loss, T = temperature (K), Z = pre-exponential factor, R = molar gas constant, q = heating rate and n = reaction order; estimated by Horovitz-Metzger method.

The thermodynamic parameters of the thermal degradation step: enthalpy (ΔH^*), entropy (ΔS^*), and Gibbs energy (ΔG^*) of activation are calculated using the following standard equations:

$$\Delta S^* = R \ln \frac{Zh}{kT_{max}}$$

$$\Delta H^* = E_a - RT_{max}$$
$$\Delta G^* = \Delta H^* - T_{max} \Delta S^*$$
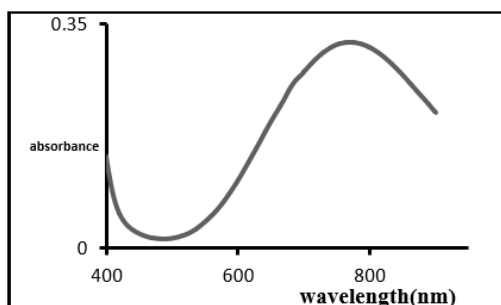
where z, k, and h are the pre-exponential factor, Boltzmann and Planck constant, respectively.

The TG and DTA thermograms of [Cu(ur)(gly)(H$_2$O)$_2$]Cl complex (Figures 8 and 9) are characterized by the three fast decomposition steps (25-318, 318-361 and 361-375 °C). The T$_{DTG}$ at 302 °C is consistent with the evolution of 100% of coordinated water, 100% of bonded chloride and 60% the urea ligand (calc. 39.95%, found 39.93%). The activation energy calculated is 89 kJ mol^{-1} (Table 6). The remaining urea molecule may be eliminated in the second step together with 52.72% of glycine molecule (calc. and found 23.43%). In this step (318-361 °C), the activation energy is 123 kJ mol^{-1} and the order of decomposition reaction is 3.6 with the apparent T$_{DTG}$ (334 °C) and the exothermic (T$_{DTA}$) peak at 349 °C (Table 7). The third step, which corresponds to 17.57% loss of glycine molecule (calc. 4.86, found 4.84%) has an activation energy of 117 kJ mol^{-1}. The final residue is CuO and 0.5C as ash [(O=21.6%gly, C=8.11%gly) (calc. and found 31.78%)]. The ΔS^*, ΔH^* and ΔG^* for these three steps are calculated (-119.3, -100.1 and -183.8 J K^{-1} mol^{-1}), (86.5, 120.2 and 114 kJ mol^{-1}) and (122.5, 153.6 and 180.7 kJ mol^{-1}), respectively.
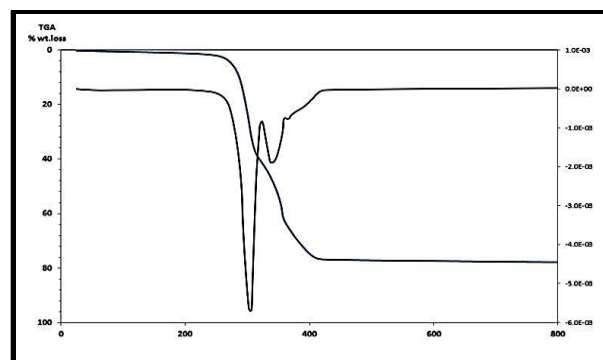
Figure 8. TG and DTG curves of [Cu(ur)(gly)(H$_2$O)$_2$]Cl.
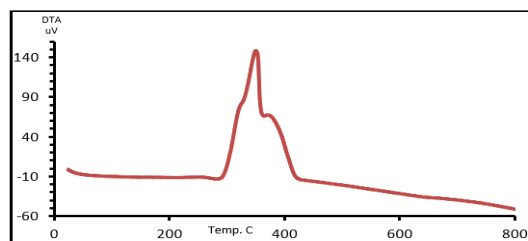
Figure 9. DTA curve of [Cu(ur)(gly)(H$_2$O)$_2$] Cl.

Table 6. Characteristic parameters of thermal decomposition (10 °C min^{-1}) for [Cu(ur)(gly)(H$_2$O)$_2$]Cl.

Comp.	Step	TGA				DTA		mass loss
		Δm% found (calc.)	T$_i$/C°	T$_f$/C°	T$_{DTG}$	T$_{DTA}$	Heat	
[Cu(ur)(gly)(H$_2$O)$_2$]Cl	1	39.93 (39.95)	25	318	3.2	323	exo	-[100%H$_2$O+ 100%Cl +60%ur]
	2	23.43 (23.43)	318	361	334	349	exo	-[40 %ur +52.72 % gly]
	3	4.86 (4.84)	361	375	363	368	exo	-[17.57% gly]
Final residue [(CuO +0.5C) (O=21.6%gly, C=8.11%gly)]: 31.78% (31.78%)								

Table 7. Kinetic and thermodynamic parameters of the thermal decomposition of [Cu(ur)(gly)(H$_2$O)$_2$]Cl.

[Cu(ur)(gly)(H$_2$O)$_2$] Cl	1	0.9918	3.6	3.7x10^6	3.2	89	-119.3	86.5	122.5
	2	0.9939	3.6	4.1x10^7	334	123	-100.1	120.2	153.6
	3	0.9862	4.9	1.9x10^3	363	117	-183.8	114	180.7

3.6. Antibacterial assay of synthesized complexes

Urea showed activity against the *Bacillus* spp. and *Escherichia coli* with inhibitory zones of 12 mm and 10 mm, respectively and glycine against the *Bacillus* with inhibitory zone of 9 mm. But no inhibition zone was observed for all the complexes against the four studied strains (*Bacillus* spp., *Escherichia coli, Pseudomonas aeruginosa* and *Staphylococcus aureus*) excepting the complex [Ni(ur)(gly)(H$_2$O)$_2$]Cl which was active against *Escherichia coli* with inhibitory zone 5 mm. This is probably because urea denatures protein when dissolved, and for the presence of amino and carbonyl groups. However, after complexes formation there would be no activity, due to the coordination of the amino and carbonyl groups[31].

4. Conclusions

The formulae and the stoichiometry of the complexes of urea and glycine with Co(II), Ni(II) and Cu(II) metal ions are suggested based on the analytical data and TGA results. Neutral bidentate behavior of the urea coordination through the amine nitrogen and carbonyl oxygen is identified by IR spectra. Glycine behaved as an anionic bidentate ligand through the carboxylate group and the neutral amino group. The electrolytic nature of the complexes was confirmed by the molar conductance values. All the complexes have an octahedral geometry, as revealed the spectral and magnetic results. The thermal decomposition studies of [Cu(ur)(gly)(H$_2$O)$_2$]Cl allowed to access the kinetic parameters for the successive steps of its decomposition. The complexes have no

antibacterial activities against the four strains of bacteria, except the Ni-complex, which is active against *Escherichia coli*, probably due to protein denaturation.

5. Acknowledgments

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Simple, fast and inexpensive method for determination of ranitidine hydrochloride based on conductometric measurements

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ABSTRACT: This work aims the development and optimization of an alternative method for ranitidine hydrochloride (RAN-HCl) determination. The proposed method was based on conductometric titration of RAN by precipitation of AgCl solid using a solution of $AgNO_3$ as titrant. It was investigated the possibility of performing the titrations on hydroalcoholic and deionized water medium. A limit of detection of 1.0 mmol L^{-1} and 0.5 mmol L^{-1} were achieved for RAN titration in deionized water and in a 75:25 hydroalcoholic mixture, respectively. Such behavior is attributed to the dielectric constant of hydroalcoholic medium, which is lower than aqueous solution, making AgCl more insoluble and improving the resolution of the conductivity curve around the end point. Therefore, it is concluded that the conductometric titration method to determine RAN using $AgNO_3$ as titrant proved to be feasible at low drug concentrations. The statistical calculations for obtained results suggested good precision for the conductometric method. According to t-test, there were no significant differences between found values at a 95% confidence level. Moreover, obtained results showed an excellent performance of the proposed method on quality control of RAN-HCl in generic formulations without any sample pretreatment.
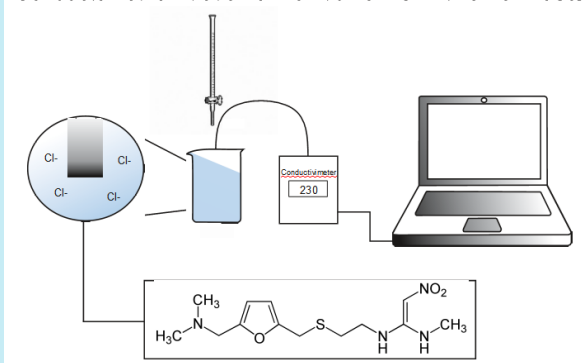
1. Introduction

In recent years, studies indicate that about 10% of the world population will suffers lesions in the gastric mucous membranes or in the duodenum[1]. The more effective treatment of this type of lesion is based on mainly administration of drugs proton pump inhibitors, as omeprazole[2]/pantoprazole[3] or anti-histamine which ranitidine hydrochloride could be highlighted[4]. Ranitidine hydrochloride (RAN-HCl - Figure 1) plays in the organism action inhibiting the secretion of gastric acid, reducing both volume and content of acid and pepsin secretion[5]. This drug is largely commercialized around the world and consequently the development of feasible, fast and sensitive methods for pharmaceutical quality control is very important.
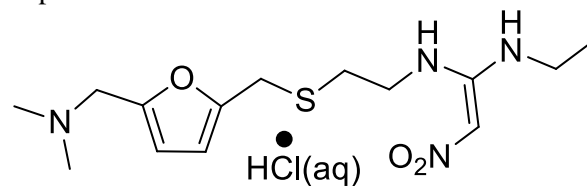

Figure 1. Molecular structure of ranitidine hydrochloride.

There are several procedures reported in literature for ranitidine determination based on chromatographic[6, 7], UV-Vis spectroscopy[8-10] and voltammetric methods[11-13]. British Pharmacopeia[14] describes the dosage of RAN-HCl in pharmaceutical formulations for quality control by liquid chromatography using potassium dihydrogen phosphate buffer and acetonitrile as mobile phase with UV detection. Brazilian Pharmacopeia[15] suggests direct molecular absorption spectrophotometry with detection at 314 nm as official method for determination of ranitidine hydrochloride. These cited methodologies are very reliable and exhibit sensibility, precision and accuracy adequate for concentration levels of pharmaceuticals formulations of RAN-HCl. However, these techniques demand long-time for sample preparation/treatment, use large volumes of organic solvents and/or require sophisticated and expensive instruments.

In order to overcome the drawbacks showed by other techniques, titrimetric procedures are an interesting alternative way for analysis of pharmaceutical formulations and/or biological samples, since this type of methodology is rather simple, with fast response and low-cost instrumentation. Moreover, detection of end point can be performed using an instrumental technique which is a practical approach to minimize analyst errors and leads more rapid, precise and accurate results[16].

Several "hydrochlorides" such as RAN-HCl can be found as active ingredients in commercialized pharmaceutical formulations. In general, hydrochlorides are more water-soluble and provide better absorption by human body[17]. Some methodologies have been described for determination of these compounds, based on indirect quantification of drug based on chloride ions released from salt dissolution[18-21]. Caetano et al.[18] reported a low-cost conductometric method for determination of verapamil hydrochloride in pharmaceutical formulations based on chloride ions released from verapamil hydrochloride which were titrated with an aqueous solution of silver nitrate. Similar approach was adopted by Noronha et al.[19] who described potentiometric and conductometric titration of diltiazem hydrochloride (DTZ) determination based on the chemical reaction between chloride ions coming from diltiazem hydrochloride molecule and Ag(I) ions, yielding the precipitate AgCl solid. The limit

of applicability of the conductometric method based on precipitation titration is defined by solubility of solid yielded AgCl (Kps ~1.8×10^{-10}). The solubility product constant can be affected by alteration in the dielectric constant of the solvent (ε) allowing to use lower concentration level of reagents. A decrease in the Kps could be obtained simply using of hydroalcoholic medium which presents a minor value of dielectric constant when compared to aqueous solution. This effect plays an important role in the precipitation titration procedure allowing the use of lower quantity of reagents.

Based on above-cited information, the present work describes a practical and accurate methodology for RAN hydrochloride determination based on conductometric titration of chloride ions using by $AgNO_3$ as titrant in hydroalcoholic medium.

2. Materials and methods

2.1. Instruments, reagents and chemicals

A system comprising by Conductometer Oakton® COM 500 and a classic burette of 25.0 mL associated with a magnetic stirrer were used for all conductometric measurements. The solutions were prepared with deionized water obtained from Milli-Q system or ethanol (Carlo Erba, 99.9%). Other used reagents were silver nitrate and sodium chloride (Merck, > 99% content) and ranitidine hydrochloride (Aldrich, > 95% content).

Silver nitrate solutions were used in the titration of the same concentration of ranitidine solutions ranging from 0.10 to 1.0×10^{-3} mol L^{-1}. The same were standardized using anhydrous sodium chloride as primary standard. For comparative method, a spectrophotometer Hewlett Packard, model 8452A equipped with 1.0 cm light path quartz cuvette. Acid-base titrations were performed using a 780 pHmeter Metrohm equipped with combined glass electrode associated a classic burette of 25.0 mL and magnetic stirrer.

2.2. Samples and standard solutions

Pharmaceutical samples in tablet form containing ranitidine hydrochloride (RAN-HCl) were purchased in local drugstore. For each sample, twenty tablets were crushed using mortar and pistil and 50.0 mL of a stock solution with theoretical concentration of 1.0 mmol L^{-1} was

prepared using deionized water or hydroalcoholic solution in different proportions of solvent.

2.3. Conductometric titration

Aliquots of 10.0 mL or 5.0 mL (reference solutions or sample) were transferred to beakers and titrated with solutions of AgNO₃ at the same concentration of the analyte (between 5.0×10^{-4} to 0.1 mol L^{-1}) with additions of 0.5 mL or 1.0 mL. Conductance values were recorded 10 seconds after the titrant addition. Before plotting titration graphs, conductance values were corrected according to volume of titrant added (Equation 1), all titrations were conducted in temperature controlled at 20 °C.

$$L_{corr} = L_{exp} * \left(\frac{(V_{ini} + V_{add})}{V_{ini}} \right) \tag{1}$$

where: L_{corr} = corrected conductance, L_{exp} = experimental conductance, and V_{ini}, V_{add} the initial volume and added volume, respectively.

From graph of corrected conductance vs. volume of AgNO₃ solution, the end point was obtained by the intersection of the straight-line segments, and thus determining the concentration of RAN-HCl.

2.4. Acid-Base titration (pH curves)

A standard solution of RAN-HCl, 0.10 mol L^{-1} was titrated with NaOH at the same concentration, previously standardized by sodium biphthalate. Based on the pH vs. NaOH volume curve was possible to estimate the pKa of RAN-HCl. The pKa value was estimated using the pH obtained at ½ volume of the end point.

2.5. Comparative method

Comparative method used is described on Brazilian Pharmacopeia[13] based on UV-Vis spectroscopy. A portion equivalent to one tablet was dissolved in water and after diluting to concentration around to 0.00125% (w/v). Standard solution was prepared at same concentration and the absorbance measurements of samples and standards solution were carry out at 314 nm using a quartz cuvette.

3. Results and discussion

3.1 Preliminary studies

Firstly, the potentiality of conductometric titration for proposed determination was investigated in order to show that indirect quantification of RAN-HCl can be realized based on precipitation reaction between Cl⁻ and Ag⁺ ions leading to formation of AgCl. An aliquot of 10.0 mL of 0.10 mol L^{-1} RAN-HCl standard solution was prepared in deionized water and it was titrated with a standardized solution of AgNO₃ at the same concentration (Figure 2). Typical conductometric curve for titration of RAN-HCl with silver nitrate shows two linear segments with significant difference in the slope. Before the endpoint, no significant variation of conductance values was observed which is attributed to consume of chloride ions by precipitation of AgCl (solubility, 1.1×10^{-5} mol L^{-1}) concomitantly with reposition of nitrate ions from AgNO₃. Slight variation in conductance observed in this segment can be attributed to exchange of ions with very similar ionic mobility (Cl⁻ 76.3 S cm² mol^{-1} and NO₃⁻ 71.5 S cm² mol^{-1}). After the endpoint, a marked increase of conductance values was observed due to excess of silver and nitrate ions in solution. The endpoint was determined by intersection of segment #1 and #2 and RAN-HCl concentration was estimated based on 1:1 stoichiometric relationship.
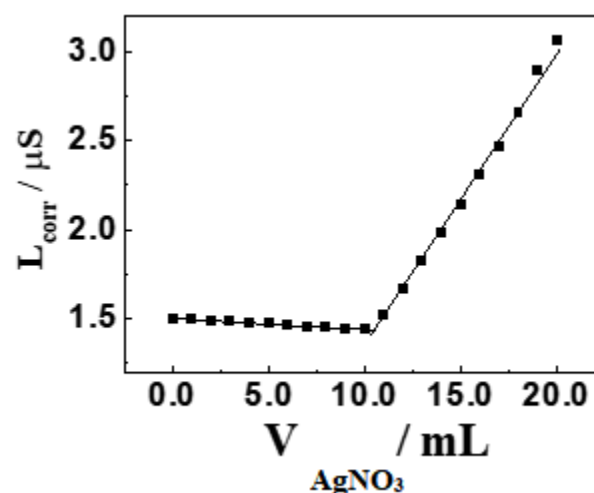

Figure 2. Representative conductometric curve obtained for titration of 10.0 mL of 0.10 mol L^{-1} RAN-HCl standard aqueous solution with a standardized solution of AgNO₃ 0.10 mol L^{-1}.

The proposed procedure is based on determination of chloride ions released from RAN-HCl after its dissolution (RAN-HCl(s) + H_2O → RAN(aq) + H_3O^+(aq) + Cl^-(aq)). Potentiometric titration of RAN-HCl with NaOH standard solution was performed in order to verify adequate reactions that occur during the conductometric titration. In Figure 3A is possible to observe a curve with similar shape to obtained titration for systems composed by weak acid titrated with strong base (endpoint is observed at pH > 7.0). Additionally, from titration curve the pKa value for reaction RAN-H^+ + H_2O ⇆ RAN + H_3O^+ was estimated as 8.05 ± 0.10 (n = 3). Thus, based on these results we believe that reactions involved in the proposed procedure could be better attributed to: RAN-HCl(s) → RAN-H^+(aq) + Cl^-(aq) (dissolution step) and Ag^+(aq) + Cl^-(aq) → AgCl(s) (titration step).

UV-vis spectrophotometric measurements were performed at different pH values in order to confirm the protonation of ranitidine molecule. Figure 3B shows UV-vis spectra where an increase in the absorption intensity at 310 nm with the increase of pH values can be observed. This behavior is attributed to protonation of RAN molecule which is majority form at pH values at least 2.0 units lower than pKa. These studies confirm that the conductometric titration was performed with RAN molecules in the protonated form.
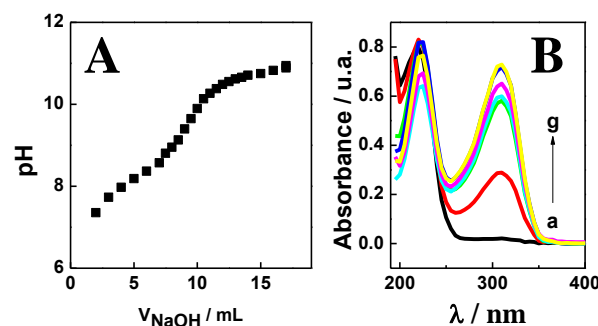

Figure 3. A) pH curve obtained for titration of RAN-HCl using NaOH standardized solution; B) Absorption spectrum in the UV-Vis region for RAN-HCl in pH values: 1.0 (a), 2.0 (b), 2.5 (c), 3.0 (d), 4.0 (e), 5.0 (f) and 6.0 (g).

3.2. Optimization of the titration method

Several investigations were realized in order to optimize the proposed methodology. For this, all solutions were prepared with deionized water to obtain a lower background signal. Firstly, the effect of solution concentration (titrant and titrated) has been evaluated by performing of three successive titrations using concentrations in range between 0.50 and 10.0 mmol L^{-1}. Table 1 presents results of recovery studies.

Table 1. Titrations realized with deionized water.

Concentration of ranitidine added / mmol L^{-1}	Concentration of ranitidine found / mmol L^{-1}	Recovery
10	9.9	99%
1.0	0.92	92%
0.50	-	ND

ND: Not detected

Adequate recoveries values were obtained using solutions with concentration of 10.0 mmol L^{-1} indicating a well-defined shape of titration curve. When concentrations of 1.0 mmol L^{-1} were used, a significant decrease in recoveries values were found suggesting a poor profile of the conductometric titration curve. Concentration lower than 1.0 mmol L^{-1} did not provide a variation in the conductance of solution which did not allow found an end point from titration curve. Thus, the best set of results using aqueous solutions were found only for concentrations higher than 1.0 mmol L^{-1}.

In order to achieve detections below the limit imposed by use of aqueous solution, we decided to try to alter the solubility of solid formed (AgCl) during titration process by using of another solvent mixed on aqueous solutions. The adopted strategy was based on variation of dielectric constant (ε) of solution. The dielectric constant of a solvent could be related with its polarity and ability for solvation of ions. When a dielectric constant of solvent is higher it means that this solvent is more polar, and it is more efficient to solvate the ions. As consequence of this, an increase in solubility of ionic solid could be observed for solvent with high

dielectric constant. The dielectric constant of water is 78.3 (25 ºC) and it is higher than ethanol (24); so, a higher solubility of AgCl solid in water is verified when compared with ethanol. In this way, the use of hydroalcoholic solutions promotes a decrease in the solubility of silver chloride and allows to use of more dilute solutions in the conductometric titration. Titrations employing hydroalcoholic solutions in the range of 0 to 75% (v/v) of ethanol were performed using 1.0 mmol L^{-1} of RAN-HCl and $AgNO_3$ in order to investigate the effect of mix of solvents (Figure 4).
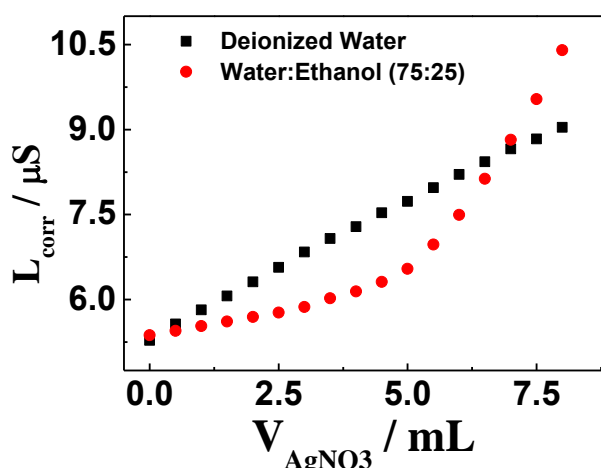

Fig. 4 Titrations conductometric curves obtained using different solvents composition.

By using of hydroalcoholic solutions, recoveries values close to 100% were obtained for all titrations, indicating that presence of ethanol as solvent results in an improvement in the determination. Amounts of ethanol higher than 50% did not yield significant differences between the segments of titration curve which can be related with decrease in the solubility of reagents. For larger amounts of alcohol (25:75 water:ethanol) there was a decrease in sensitivity caused by a poor solubility of $AgNO_3$. Using a composition of 75:25 water:ethanol, it was possible to determine the concentration of RAN-HCl employing concentration of 0.5 mmol L^{-1} for reagent which was not achieved performing the titration in water. Such behavior was attributed to the lower dielectric constant inherent to the solution which resulted in a lower background signal enabling the identification of titration equivalence point with good reliability.

3.3. Determination of RAN-HCl in pharmaceutical samples

After the optimization of experimental conditions, the proposed method was applied for determination of RAN-HCl in triplicate for three different samples of pharmaceutical formulations (Table 2). Results obtained using the conductometric titration were compared with those provided using the spectrophotometric method recommended by the Brazilian Pharmacopoeia and the values given on the pharmaceutical formulations labels.

Table 2. Results of determination of RAN-HCl in pharmaceutical formulations.

	Sample	Labeled values / mg	Proposed Method / mg	Reference Method / mg	RSD*
1	Generic	300	305 ± 9	298.4 ± 0.7	2.1%
2	Generic	150	158.7 ± 0.3	165 ± 6	3.5%
3	Generic	150	161.1 ± 0.2	163 ± 5	1.0%

*RSD: relative standard deviation (Proposed *vs.* Reference Method)

Statistical test suggested good accuracy for the conductometric method in comparison with official methodology, indicating that the excipients used to manufacture the tablets of ranitidine formulations did not exhibit interference in the determination of the drug[18-20]. According to the t-test, there were no significant differences between the values found at the 95% confidence level. Moreover, results obtained showed an excellent performance of proposed method to quality control of RAN-HCl in generic formulations without any sample pretreatment.

4. Conclusions

This study demonstrated that conductometric titration is a very powerful methodology for determination of ranitidine hydrochloride, being a simple, fast and inexpensive alternative route to perform quality control of these species, especially in small laboratories. The use of hydroalcoholic solutions allowed a decrease in the detection limit of the technique. It was possible to perform quality control of RAN-HCl in the order of 0.5 mmol L^{-1} with significant reducing costs. Pharmaceutical samples without any pretreatment were analyzed using proposed titration and values with excellent agreement with an official method were found.

5. Acknowledgments

6. References

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
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Dark breather using symmetric Morse, solvent and external potentials for DNA breathing

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ABSTRACT: We analyze the dynamics and the quantum thermodynamics of DNA in Symmetric-Peyrard-Bishop-Dauxois model (S-PBD) with solvent and external potentials and describe the transient conformational fluctuations using dark breather and the ground state wave function of the associate Schrodinger differential equation. We used the S-PBD, the Floquet theory, quantum thermodynamic and finite difference methods. We show that for lower coupling dark breather is present. We estimate the fluctuations or breathing of DNA. For the S-PBD model we have the stability of dark breather for k<0.004 and mobile breathers with coupling k=0.004. The fluctuations of the dark breather in the S-PBD model is approximately zero with the quantum thermodynamics. The viscous and external potential effect is direct proportional to hydrogen bond stretching.
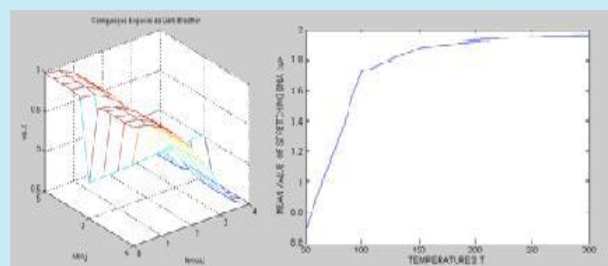
1. Introduction

The deoxyribonucleic acid DNA is a thread-like chain of nucleotides carrying the genetic information of all organisms. The coding sequences for genes and regulatory information are located in DNA and is marginally stable and undergoes a "melting phase transition". There are many experimental ways to study the fluctuations or breathing of DNA: hydrogen exchange, formaldehyde probing, protein-nucleic acid interactions, DNA replication, DNA base analogue spectroscopy, single molecule DNA-protein interactions, two-dimensional fluorescence spectroscopy[1]. The interaction between the viscous potential and external forces prevent DNA to unzip perfectly but allows DNA to split at a certain distance from its original position[2]. S. Flach gives the theory of the "discrete breathers" and applications[3].

R. S. Mackay investigates the Peyrard Bishop model for the study of nonlinear excitations travelling along the DNA chains[4]. J. Cuevas has results about breathing of DNA using the spatially localized oscillations or 'discrete breathers'[5]. The mobility and breathing of DNA depends on the harmonic bifurcation[6]. The strong dependence on sequence, temperature and salt concentration for the breathing dynamics of DNA found here points

at a good potential for applications and the effect of the viscous and external forces[2, 7].

First, the PBD model is introduced. It is then followed by the dynamical and the thermodynamic formulations. We show that mobile breather can lead to the observed breathing, but the amplitude of the breather is determinant for the transient conformational fluctuations of DNA. The results obtained in our simulation verify the existence of dark breather with the conditions describes by R.S. Mackay. The symmetric potential does not give a solution for the transition of the DNA. For that reason, it is necessary to investigate the effect of the solvent and external potentials. The calculation of hydrogen bond stretching using transfer integral operator and difference finite methods are presented.

2. Materials and methods

2.1 Symmetric Morse Potential in the PBD model

The biomechanics of DNA is represented by two degree of freedom X_n and Y_n which correspond to the displacement of the base pair from their equilibrium position along the direction of the hydrogen bonds connecting the two-base pair of nucleotides.

The studies of the Symmetric Peyrard-Bishop-Dauxois (S-PBD) models that included the modified Morse potential was done by adding the absolute value:

$$V(u) = \frac{1}{2}\left[\exp(-|u|) - 1\right]^2 \tag{1}$$

where V = symmetric Morse Potential.

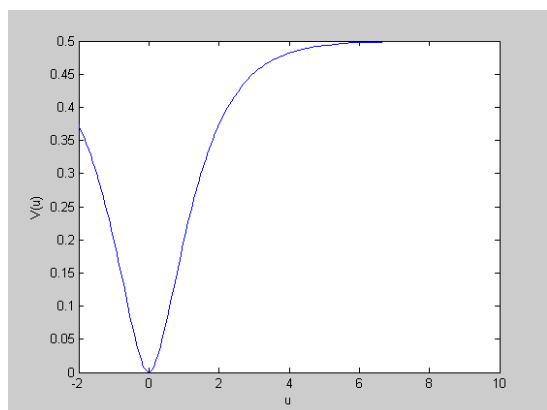The profile of symmetric Morse potential can be seen in Figure 1.

Figure 1. The symmetric Morse potential.

$$H = \sum_{n=1}^{N}\frac{m}{2}\dot{u}_n^2 + \sum_{n=1}^{N}\frac{K}{2}\left(u_n - u_{n-1}\right)^2 + \sum_{n=1}^{N}V_n(u_n) \tag{2}$$

where N = number of the pairs of bases; K = *velocity* = *u* coupling constant; and u_n = stretching of the hydrogen bonds = $(X_n - Y_n)/\sqrt{2}$.

2.2 Dynamics of S-PBD

The associated equations for Equation 4 are the system equations (n=1, 2... N):

$$\ddot{u}_n + sign(u_n)\left[e^{-2/u_n\prime} - e^{-/u_n\prime}\right] + K(2u_n - u_{n-1} - u_{n+1}) = 0 \tag{3}$$

Using the approximation for the oscillator n and $T = 2\pi/w_b$

$$u_n = z_n^0 + \sum_{k=1}^{k_m} 2z_n^k \cos(k\omega_b t) \tag{4}$$

and substituting in Eq. 3 one has:

$$k^2\omega_b^2 z_n^2 + V_n\prime^k + K(2z_n^k - z_{n-1}^k - z_{n+1}^k) = 0 \tag{5}$$

which depends on the parameter K, and $V_n\prime^k$ is the kth Fourier coefficient for the periodic Function $V'(u_n(t))$.

Remark 1. <u>One site dark breather</u>

The dark breather solution is obtained in conditions (t=0) where all the oscillators are at rest, but equally shifted from their equilibrium position, while the central one is at the rest. The codification for one site dark breather is 1,1,…1,0,1,…,1,1.

In Fig. 2 the dark breather is depicted. This figure shows the numerical solution of equation (5). The second derivative of the symmetric Morse potential is given by:

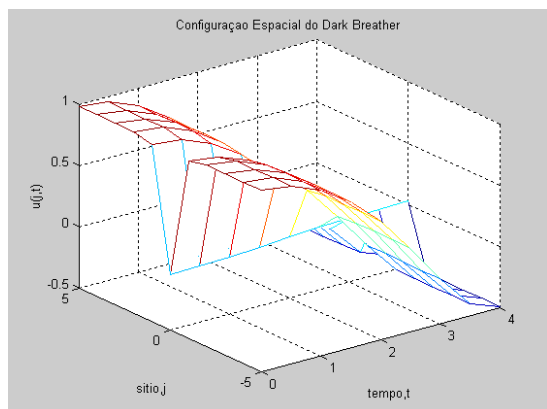$$V'' = \left[2e^{-2/u_n\prime} - e^{-/u_n\prime}\right].$$

Figure 2. Spatial dark breather configuration of the symmetric Morse potential.

The dynamics of the DNA is a set of coupled oscillators, and the vibrational motion is equivalent to equation (3) which depends of the Symmetric Morse potential and constant K of coupling.

The amplitude of the breather is determinant for the transient conformational fluctuations of DNA. In our case the Figure 2 give a small amplitude.

2.2.1 Existence of Harmonic bifurcation

We analyze the stability of the breather solution. Let us introduce a function $\tilde{u}_n(t) = u_n(t) + \varepsilon_n(t)$, where $u_n(t)$ is the periodic breather solution shown in Figure 2. The term $\varepsilon_n(t)$ is a perturbation: $\tilde{u}_n(t)$ must satisfy the system (3) and expanding around $u_n(t)$ to first order (linearization), we obtain the following system of equations for $\varepsilon_n(t)$

$$\ddot{\varepsilon}_n + \left(V''(u_n(t))\right)\varepsilon_n + K(2\varepsilon_n - \varepsilon_{n-1} - \varepsilon_{n+1}) = 0 \quad (6)$$

We can associate a monodrama my matrix for this equation with Fouquet multipliers[5].

The solution is stable if the modules of Fouquet multipliers are one. The especial instability ("harmonic bifurcation") in our case happens when a pair of Fouquet multipliers merges at $\lambda = 1$ and splits off circle onto the positive real axis in Fig. 3.
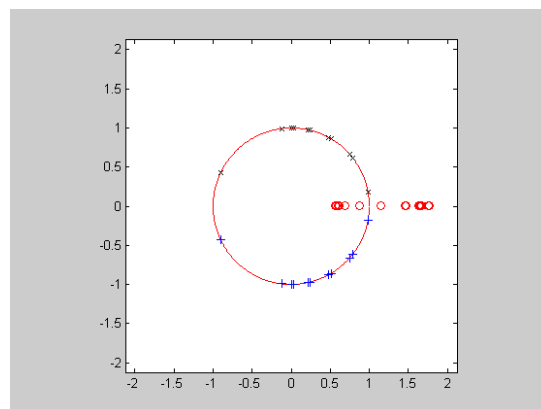

Figure 3. The instability "harmonic bifurcation" with the evolution of the Fouquet multipliers. Case SPBD model with the parameters: K=0.004, w_b=0.8 for the dark breather.

2.2.2 Existence of Mobile breather

For the coupling K=0.004 and w_b =0.8 there is a harmonic bifurcation. In this case we can construct a dark breather mobile. Once the system of equations (3) is worked out by Runge Kutta method for the Cauchy problem with the equations (3). We can use the Figure 2 for the initial conditions of the position and average speed of each position "n" respect to the harmonic oscillation corresponding to the DNA.

The center of energy of the breather mobile is given by[5]

$$X_E = \sum_{n=1}^{N} n H_n^d \ / H \quad (7)$$

where the density energy has the form

$$H_n^d = \frac{1}{2}\dot{u}_n^2 + \frac{K}{4}\left(u_n - u_{n-1}\right)^2 + \frac{K}{4}\left(u_{n+1} - u_n\right)^2 + V_n(\sqrt{2}u_n) \quad (8)$$

It is very important the initial velocity of the BM for the displacement a long of sites of DNA and can be produced of DNA breathing.

This parameter initial velocity $v(0)$ is transcendental for DNA breathing.

Remark 2. Initial velocity and the perturbation velocity

We can use the profiles of the stationary dark breather obtained from equations (3). The velocity is a vector which the components are given by

$$v_n(0) = (u_{n+1}(0) - u_{n-1}(0))/2 \quad (9)$$

We can define a perturbation velocity in terms of the parameter λ. The components of this vector perturbation *V* are given by

$$V_n(0) = \lambda(v_n(0)/v) \tag{10}$$

Where *v* is the norm of the vector of the components $v_n(0)$.

Remark 3. The <u>Cauchy problem</u> is given by

$$\ddot{u}_n + V'(u_n) + K(2u_n - u_{n-1} - u_{n+1}) = 0 \tag{11}$$

Initial conditions: $u_{.}(0)$ = profiles of the solutions of the Figure 2. The velocities are given by the expression (10) with λ = 0.1. We can obtain the solutions of the equations using initial condition with the software Fortran (for a review, see ref. 9).

2.3 Quantum Thermodynamics of S-PBD

The evaluation of the partition of equation (2) using the transfer integral operator method in the thermodynamic limit reduces to solving the pseudo-Schrodinger equation (12):

$$\{-1/(2\beta^2 K)d^2/du_n^2 + U(u_n, \beta)\}\psi(u_n) = \varepsilon\psi(u_n) \tag{12}$$

$$U(u_n, \beta) = V(\sqrt{2}u_n) + (1/2\beta)\ln(\beta K/2\pi) \tag{13}$$

$$\beta = 1/(K_B T) \tag{14}$$

We use the symmetric Morse potential. The fluctuations or breathing of DNA can be performed numerically using the finite difference methods. Firstly, we obtain the ground state wave function of equation (12). For estimate the mean value of the fluctuations we use the formula:

$$<u> = \int_{-\infty}^{+\infty} \psi^2_0 u\,du \tag{15}$$

The ground state wave function for the symmetric Morse potential is symmetric in consequence the mean value of the fluctuations is approximately zero (for a review, see ref. 12).

In Figure 4 is depicted the example of the ground state wave function for the symmetric Morse potential.
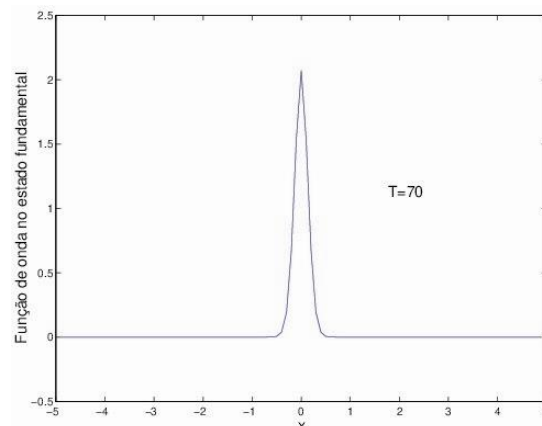

Figure 4. Ground state wave function for the symmetric Morse potential with the control parameter Temperature=70 °K.

2.3.1 Quantum Thermodynamics of S-PBD with solvent and external potentials

We can consider the new potential for the equation (13):

$$U(u_n) = V(\sqrt{2}u_n) + Vsolvent(u_n) + V_0$$
$$\exp(-0.1u_n^2) + (1/2\beta)\ln(\beta K/2\pi) \tag{16}$$

The solvent potential is given by: $V_{solvent} = 0.04*v*\tanh(u_n/5 - 1)$. In Figure 5, it is depicted the example of the solvent potential.
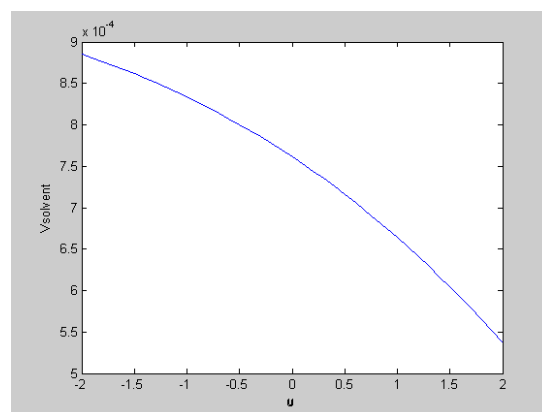

Figure 5. The solvent potential with the control parameter v=0.025.

For the symmetric Morse potential in the S-PBD Model we can get many values of the melting temperatures. For example, for T =270 K and the control parameter v = 0.001, V_0 = 0.005 the mean value of the fluctuations $<u>$ = 1.9586 Å. The hydrogen bond stretching as a function of

temperature gives a melting temperature depicted in Figure 6.

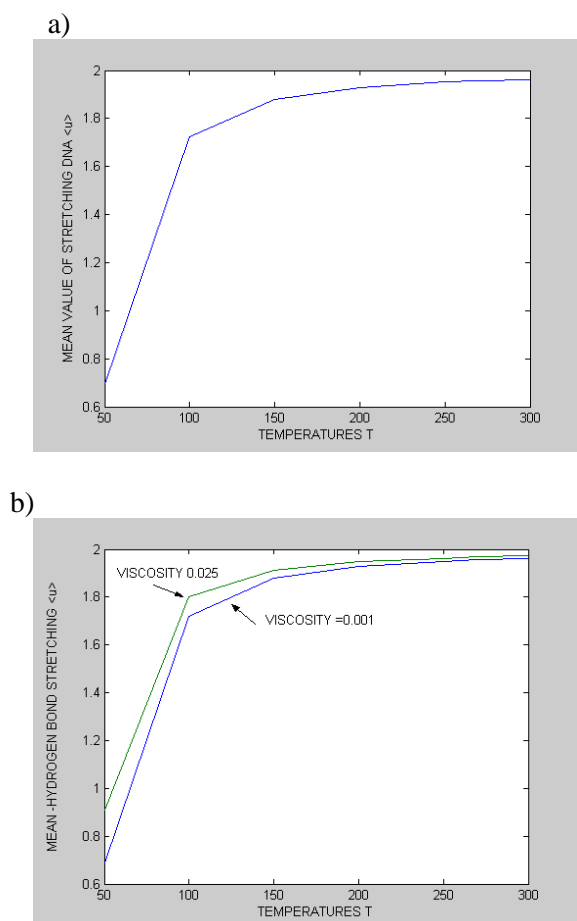Bustamante[13] has the interplay between the "DNA breathing" with the viscosity coefficient of the medium.

a)

b)

Figure 6. The hydrogen bond stretching as a function of temperature for external.

Potential $V = V_0 \ exp \ (-0.1 \ u^2 \ )$, $V_0 = 0.005$ and solvent potential with the viscosity control parameter v: a) v=0.001 and b) v=0.025.

3. Results and discussion

The solutions of the dynamical equations (5) give the dark breather mobile. We have the mobile breather using the center of energy for the initial velocity of 0.1. This method is based on the literature[6, 8].

We have obtained harmonic bifurcation using the symmetric Morse potential with the parameter K=0.004.

We have obtained the Eigen functions of the pseudo-Schrodinger equation (12) to demonstrate that the mean value breathing of DNA is zero. The analysis is based on the reference[12].

For the symmetric Morse potential in the S-PBD Model, we can get the melting temperature for T = 270 K, control viscosity parameter v = 0.001 and the constant of the external potential V_0 = 0.005. For these values the mean value of the fluctuations $<u>$ is 1.9586 Å. In this case, we can get the DNA breathing with the variations of temperatures (Figure 6a).

Figure 6b indicates that mean value of stretching $<u>$ is direct proportional to the coefficient of viscosity. The increase of the viscosity will increase the hydrogen bond stretching. The viscous and external potential effect is direct proportional to hydrogen bond stretching. For $V_0 = 0.5$ the mean value of hydrogen bond is $<u>$= 3.82 with the temperature T = 270 K and viscosity v = 0.025.

The Figure 6 shows that for T > 150 K the viscous force is not important for the DNA breathing. This result is similar to that obtained in the literature[2].

4. Conclusions

The stability of dark breathers using of the symmetric Morse potential have been obtained with the Floquet's theory. It is very important to emphasize that dark breathers at low coupling are shown to be stable in the PBD model with k<0.004. For k=0.004 we have harmonic bifurcation and the mobile dark breather. In this case and using numerical simulations we can demonstrate that the mean value of the hydrogen bond stretching is zero.

For the symmetric potentials we have significant fluctuations in the analysis of the breathing DNA with solvent and external potentials. The external potential is more important than the viscous force for the estimated melting temperature and the mean value of the hydrogen bond stretching.

5. Acknowledgments

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